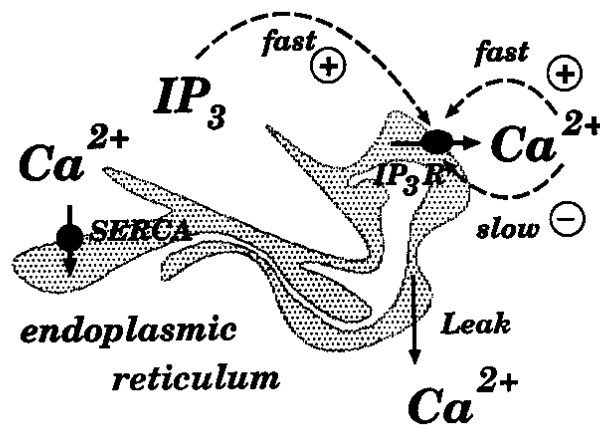


# Stochastic Modeling of Local and Global Intracellular Calcium Dynamics



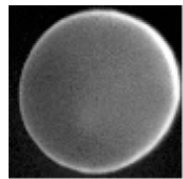
Gregory D. Smith  
Department of Applied Science  
College of William and Mary

CBL



>> Computational Biology Laboratory  
Department of Applied Science @ The College of William and Mary





1.2 mm  
(Nuccitelli)

# IP<sub>3</sub>Rs are clustered in *Xenopus laevis* oocytes

blip

fundamental

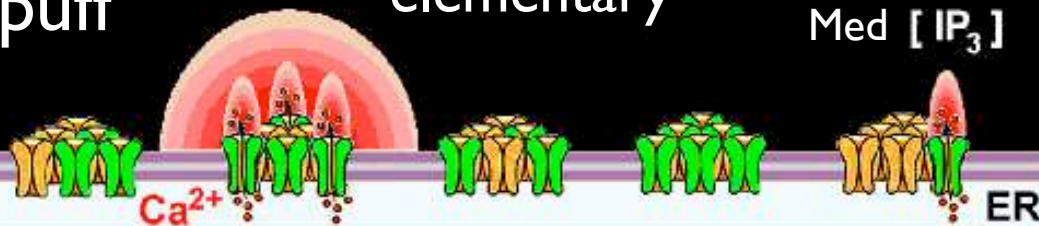
Low [IP<sub>3</sub>]



puff

elementary

Med [IP<sub>3</sub>]



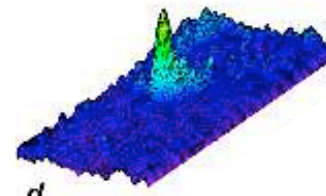
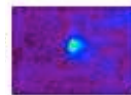
wave

global

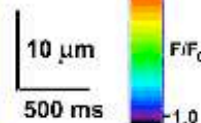
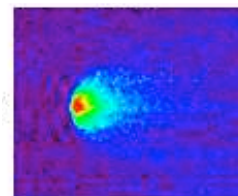
High [IP<sub>3</sub>]



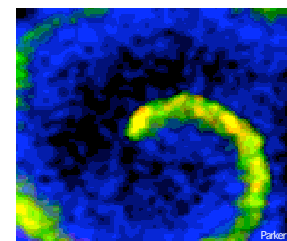
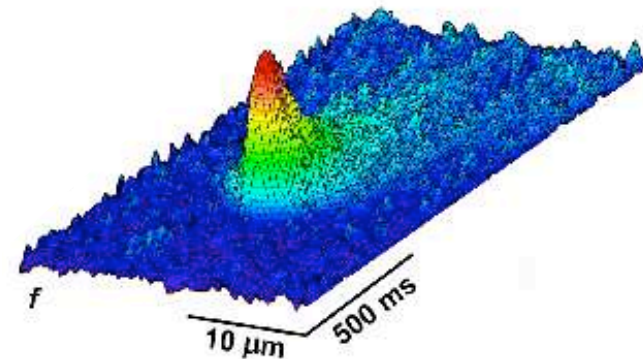
Three “modes” of Ca<sup>2+</sup> release  
(Parker, Bootman, Berridge)



blip



puff

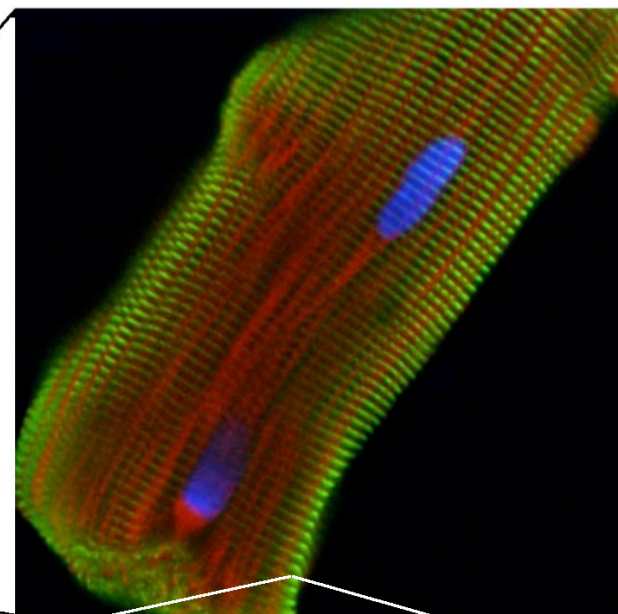
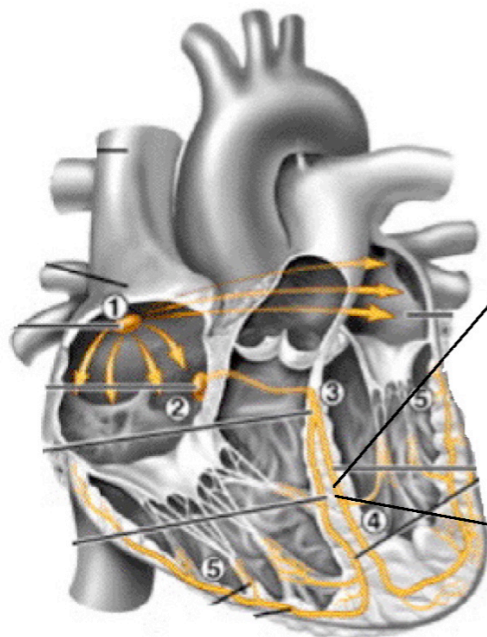


wave

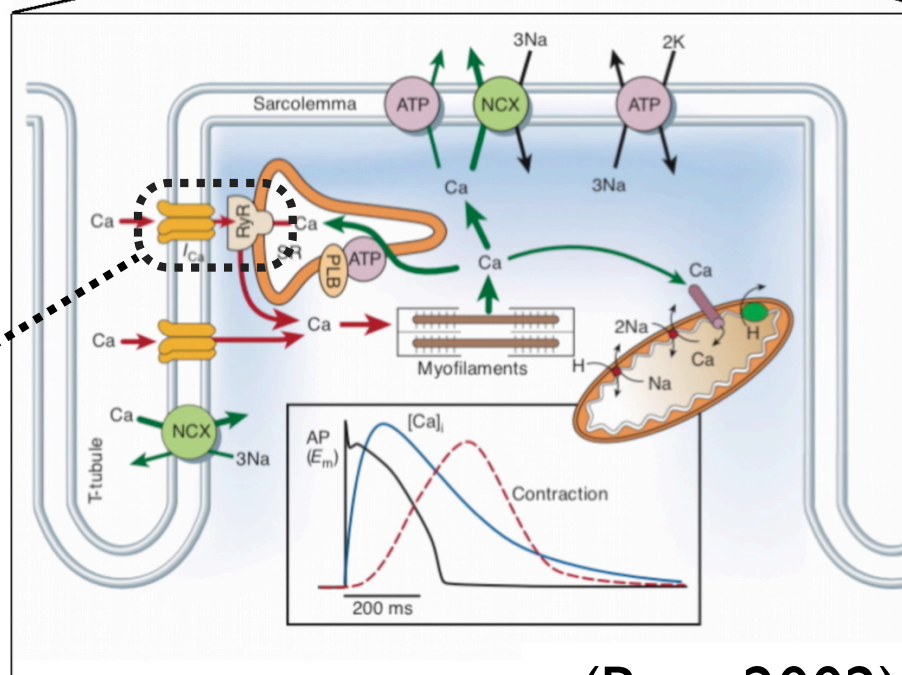
(Parker)

# RyRs are clustered in cardiac myocytes (quarks, sparks, waves)

- Depolarization
- Ca influx (DHPRs)
- Ca release (RyRs)
- Elevated Ca in myoplasm
- Contraction of sarcomere



“Common pool” models of excitation-contraction coupling do not properly account for local Ca signaling in the diadic space and junctional SR



(Bers 2002)

?

single channel gating  $\Leftrightarrow$  calcium puffs/sparks

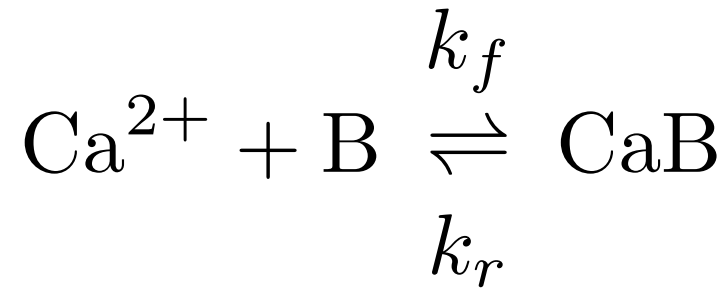
~~allosteric coupling~~

arrangement of channels

buffered calcium diffusion

# buffered Ca diffusion

near point source  
for free Ca



$$\frac{\partial[\text{Ca}^{2+}]}{\partial t} = D_{\text{Ca}} \nabla^2 [\text{Ca}^{2+}] - k_f [\text{Ca}^{2+}][\text{B}] + k_r [\text{CaB}] + \sigma(t) \delta(r)$$

$$\frac{\partial[\text{B}]}{\partial t} = D_{\text{B}} \nabla^2 [\text{B}] - k_f [\text{Ca}^{2+}][\text{B}] + k_r [\text{CaB}]$$

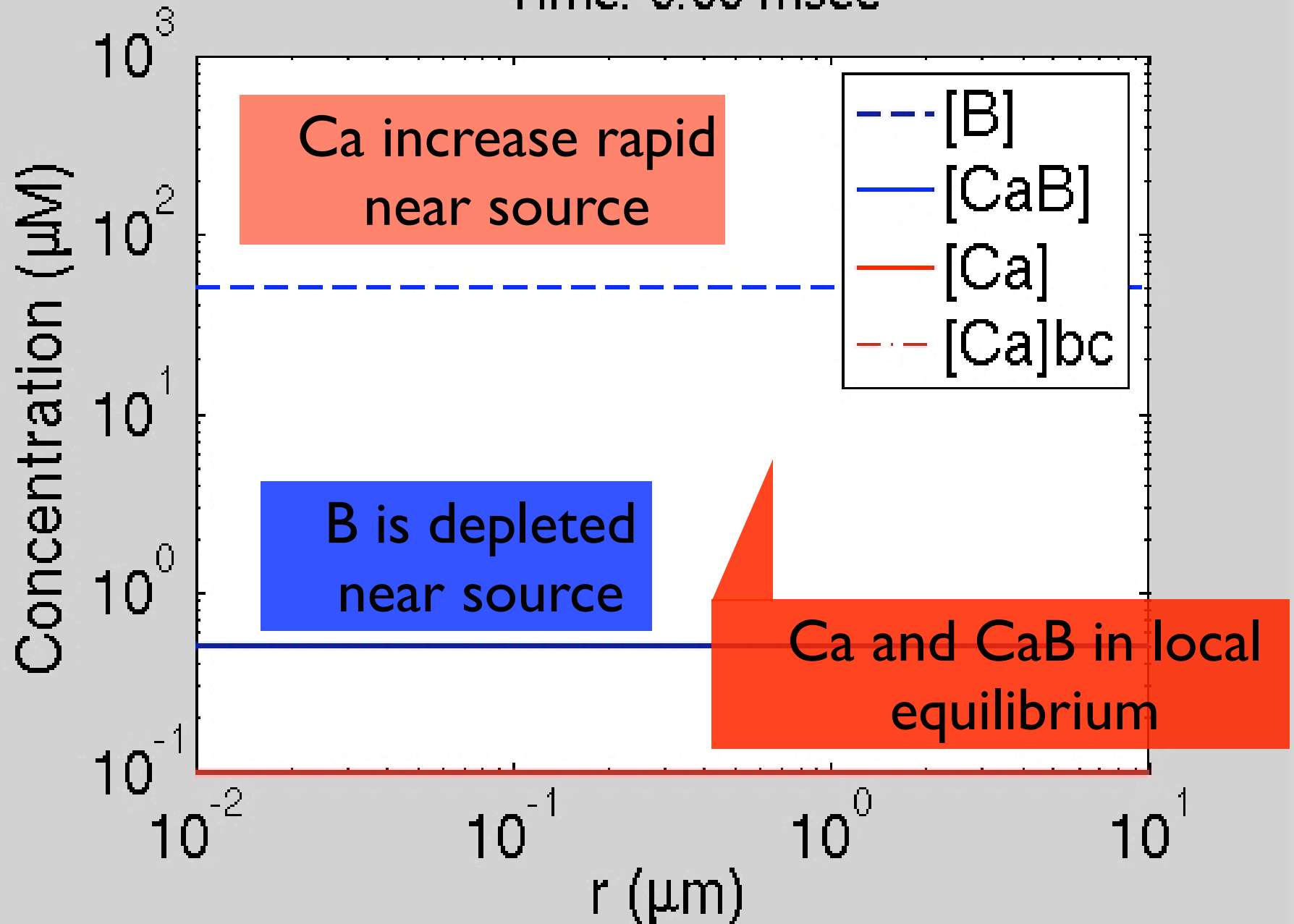
$$\frac{\partial[\text{CaB}]}{\partial t} = D_{\text{CaB}} \nabla^2 [\text{CaB}] + k_f [\text{Ca}^{2+}][\text{B}] - k_r [\text{CaB}]$$

$r = 0$  : flux for Ca turns “on” at  $t = 0$  (no flux for buffer)

$r \rightarrow \infty$  : buffers in equilibrium with background Ca

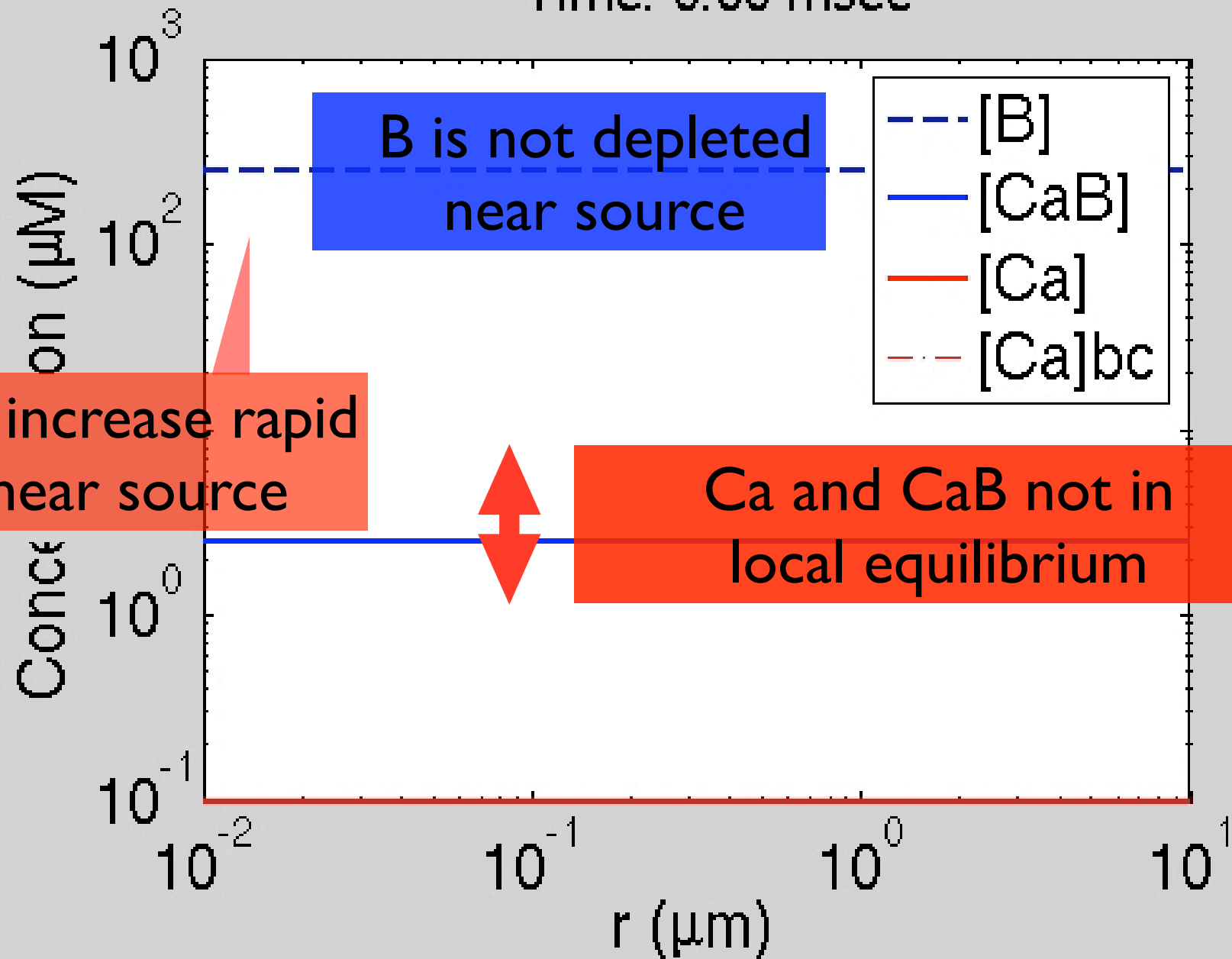
# 5 pA source — cluster of channels — fast buffer kinetics

Time: 0.00 msec



# 0.5 pA source — single channel — slow buffer kinetics

Time: 0.00 msec



Ca increase rapid near source

B is not depleted near source

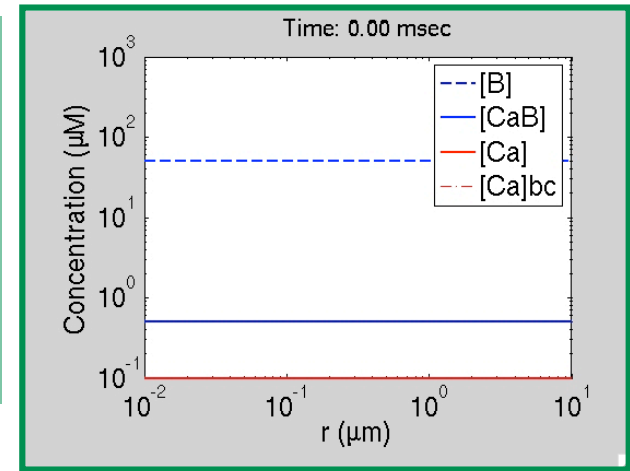
Ca and CaB not in local equilibrium

- Steady-state rapid buffer limit (large source)

$t \rightarrow \infty$

$$D_{Ca}[Ca^{2+}] + D_{CaB} \frac{[Ca^{2+}][B]_T}{[Ca^{2+}] + K} = \frac{\sigma}{2\pi r} + const$$

$$K = k_r/k_f \quad [B]_T = [B] + [CaB]$$



Buffer is nearly saturated near source

Local equilibrium between  $Ca^{2+}$  and buffer everywhere

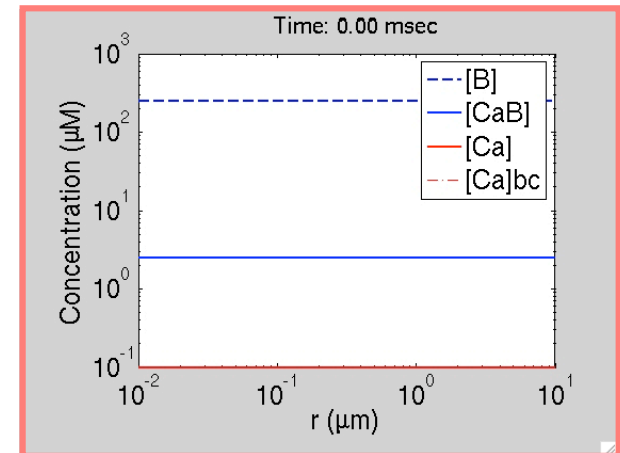
(Wagner & Keizer 1994)  
(Smith 1996)

- Steady-state excess buffer limit (small source)

$t \rightarrow \infty$

$$[Ca^{2+}] = \frac{\sigma}{2\pi D_{Ca}r} e^{-r/\lambda} + [Ca^{2+}]_\infty$$

$$\lambda = \sqrt{D_{Ca}/k_f[B]_\infty} \quad [B]_\infty = \frac{K[B]_T}{[Ca^{2+}]_\infty + K}$$



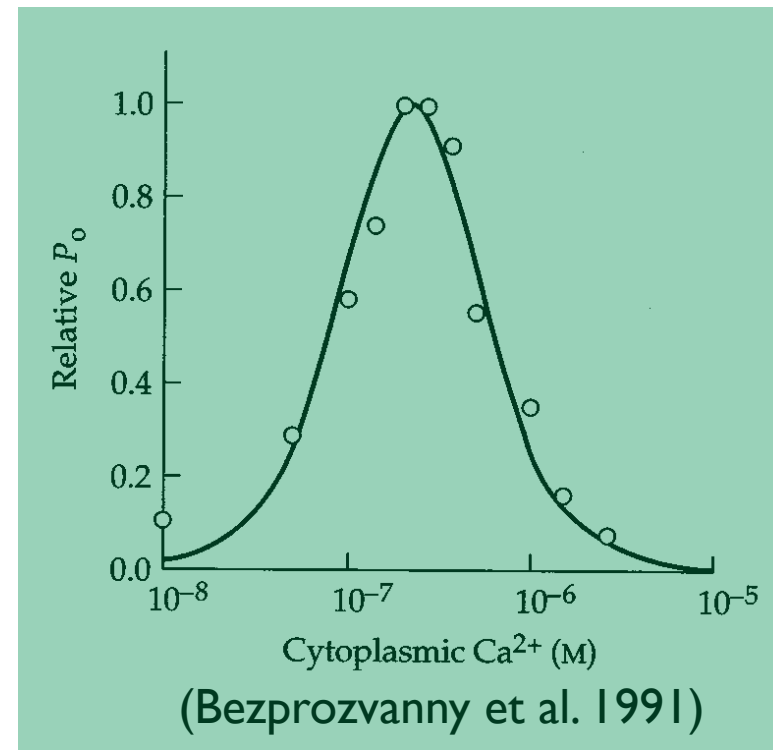
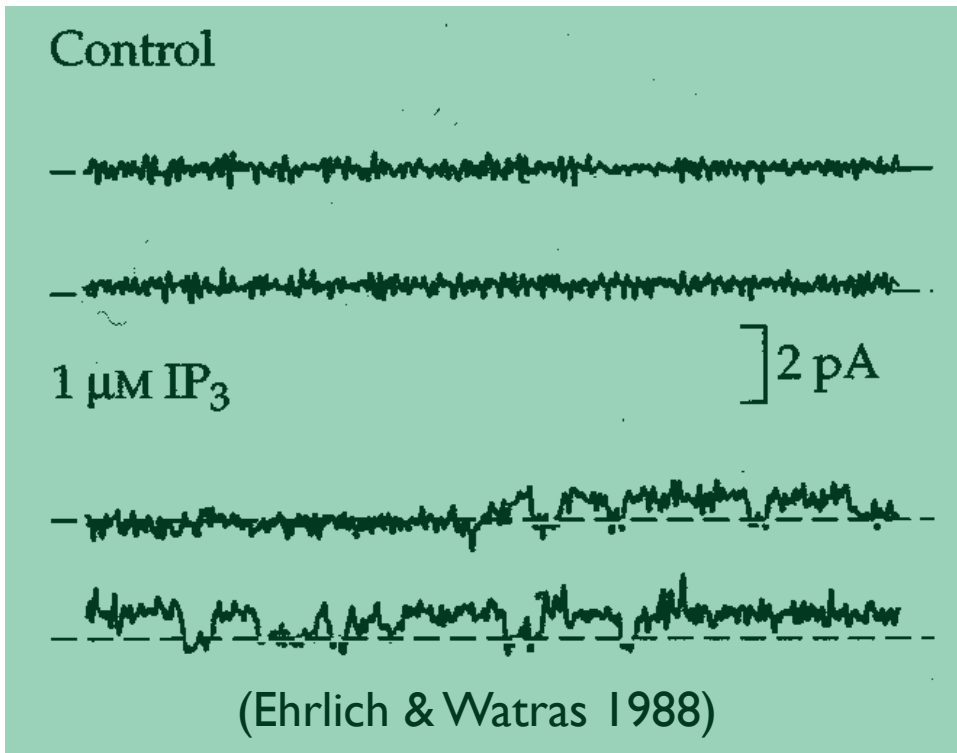
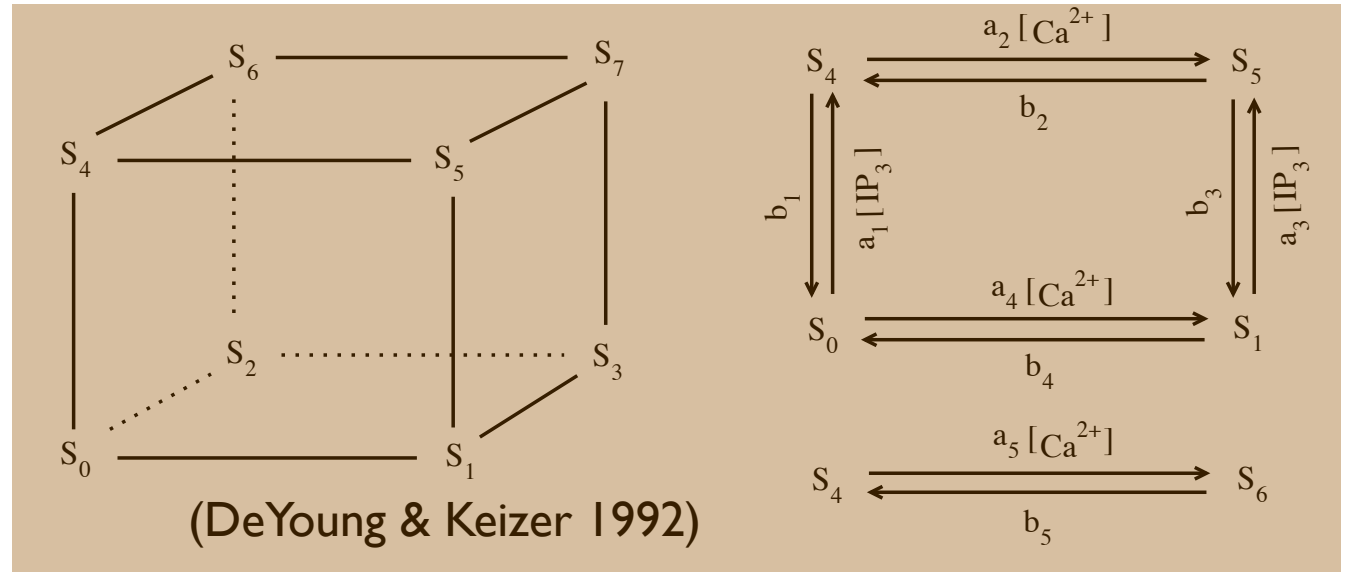
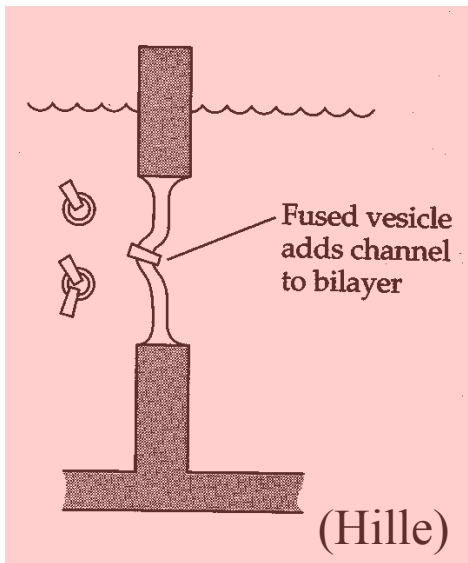
Buffer profile is only slightly perturbed near source

No local equilibrium between  $Ca^{2+}$  and buffer near source

(Neher 1986)



# IP3R and RyR gating modeled as a Markov chain



intracellular channels are modeled as Markov chains

$$X(t) \in \{1, 2, \dots, M - 1, M\}$$

each state is “open” or “closed”

$$Q(t) = K_- + c(t)^\eta K_+$$

infinitesimal  
generator

dissociation  
rate constants

local [Ca]

association  
rate constants

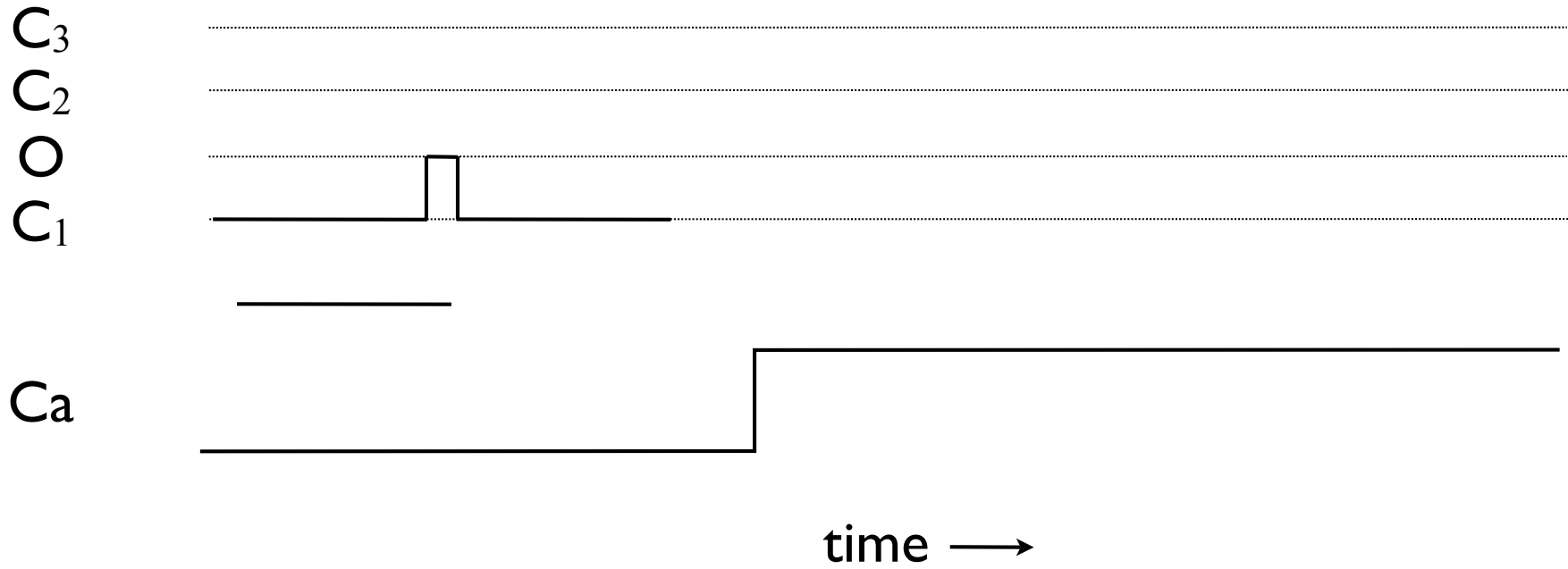
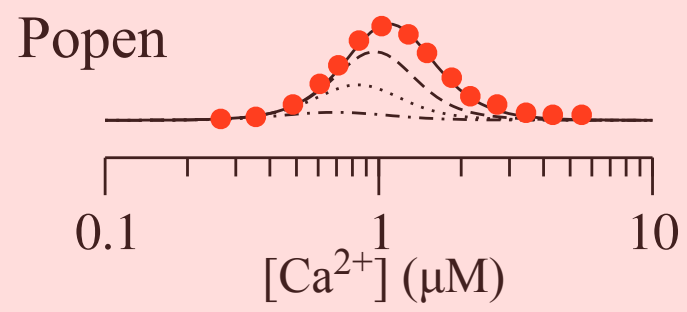
cooperativity of  
Ca<sup>2+</sup> binding

improve  
next slide

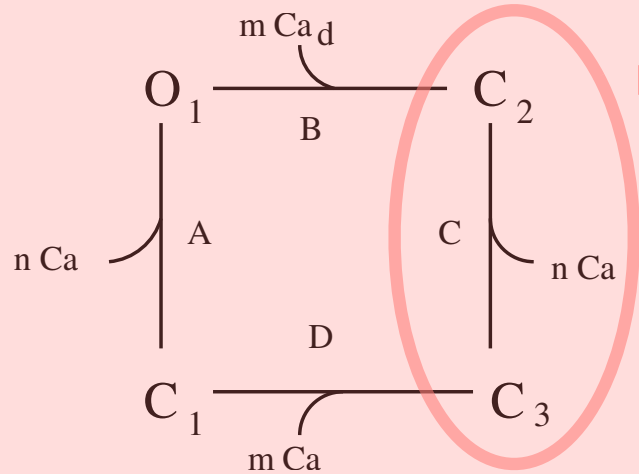
# four-state model with fast Ca activation and slow Ca inactivation



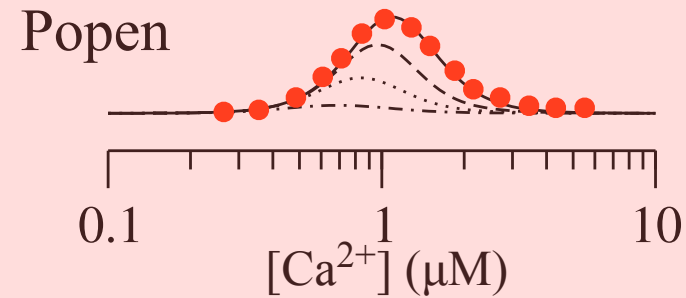
refractory states



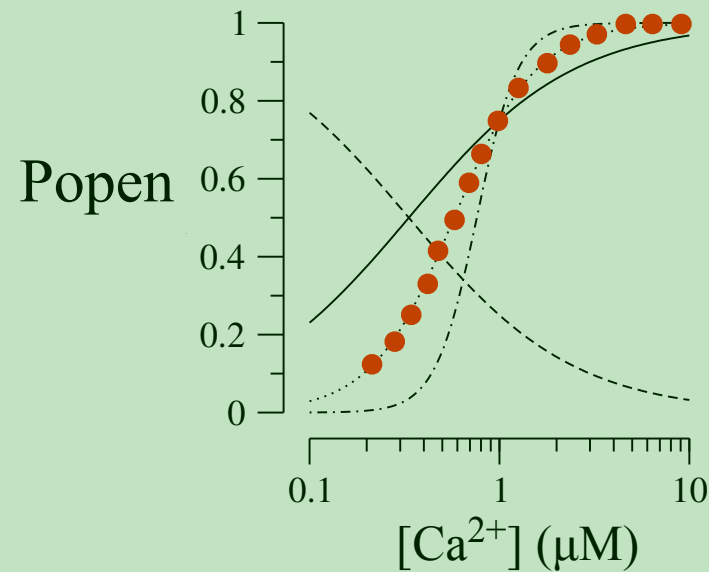
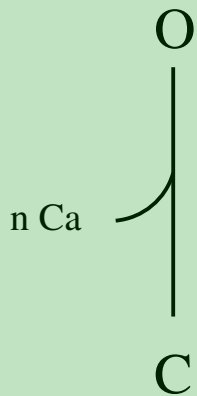
# four-state model with fast Ca activation and slow Ca inactivation



refractory states



# two-state model with Ca activation but no inactivation



channels are coupled assuming excess buffer limit

$$C = (c_{ij})$$

$N \times N$  coupling matrix

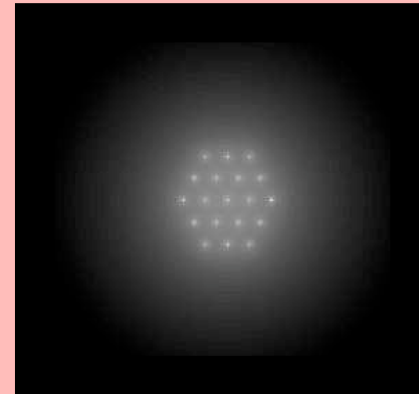
$$c_{ij} = \frac{\sigma_0}{2\pi D r_{ij}} e^{-r_{ij}/\lambda}$$

superpose interactions

instantaneous  
coupling

$$\tau_{chan} \ll \tau_{diff}$$

arrangement of channels

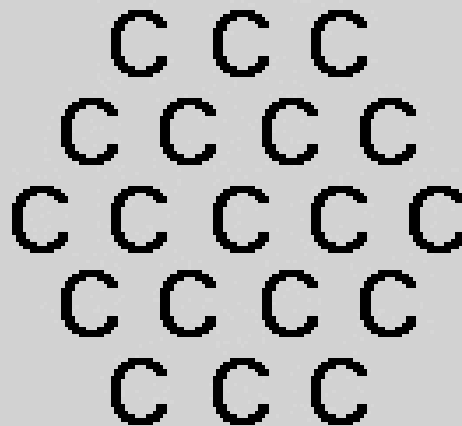
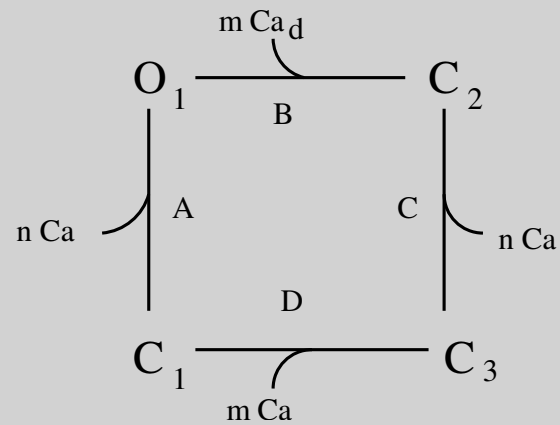
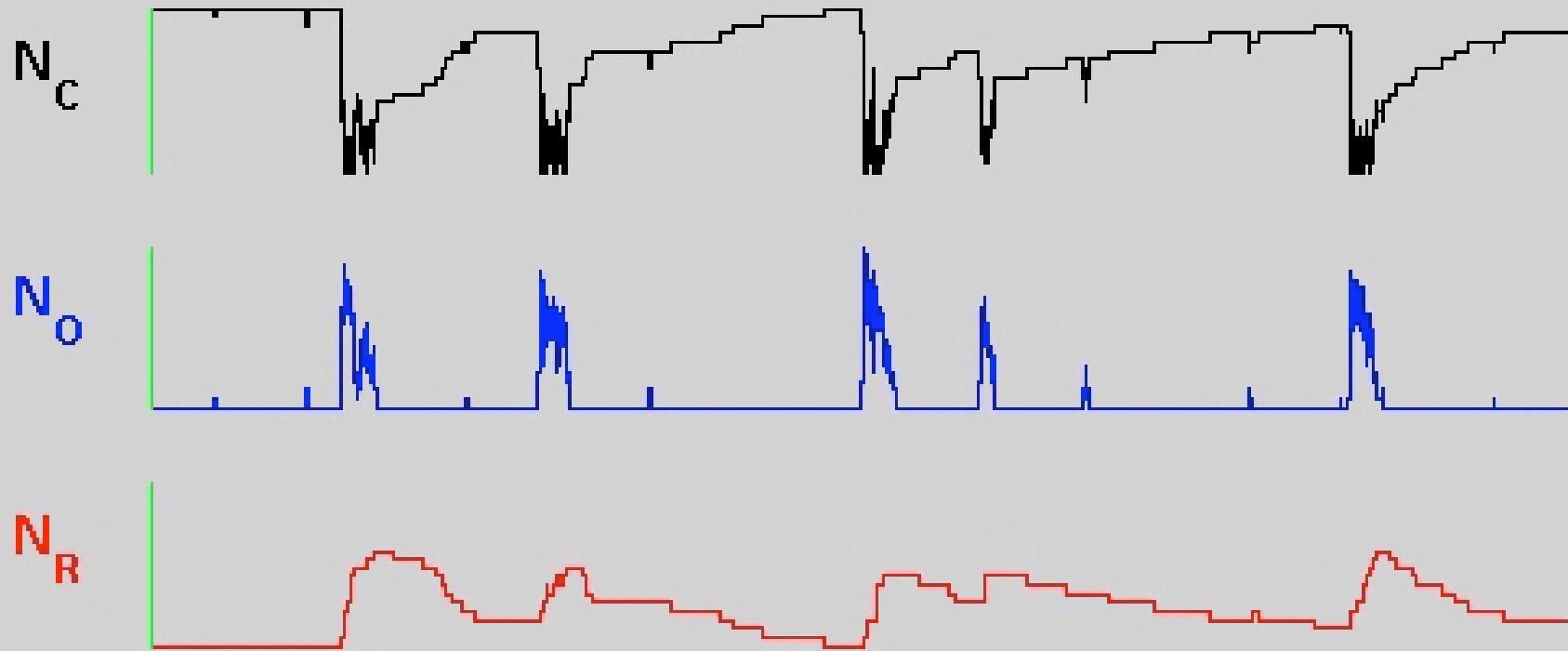


# Four-state model with $\text{Ca}^{2+}$ activation and $\text{Ca}^{2+}$ inactivation

$R = 0.12 \lambda$

Score = 0.47

Time = 0 ms

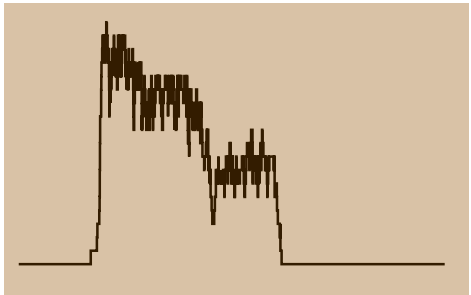
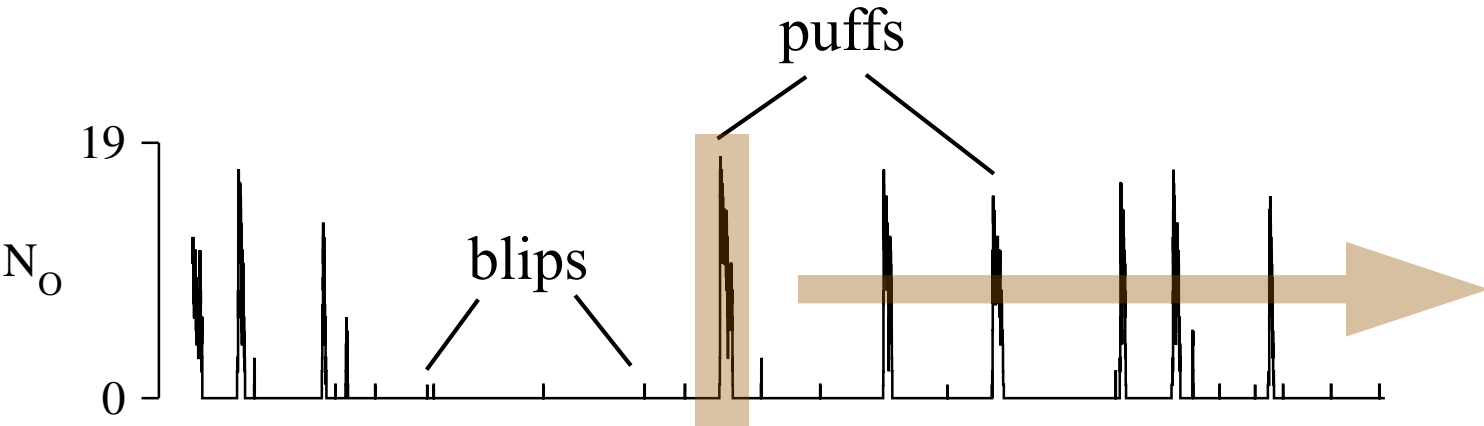
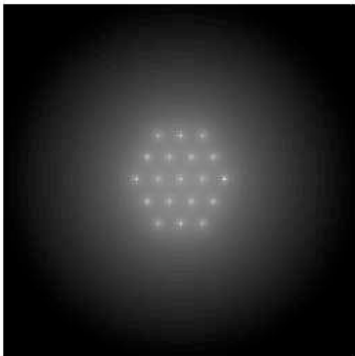
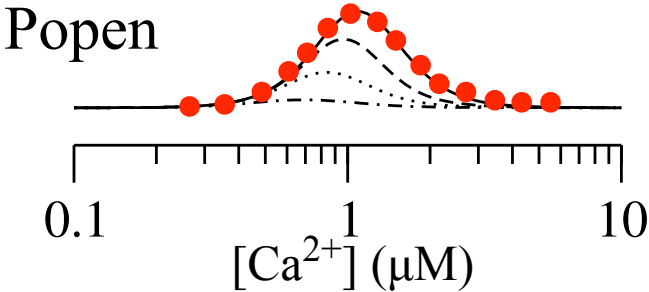
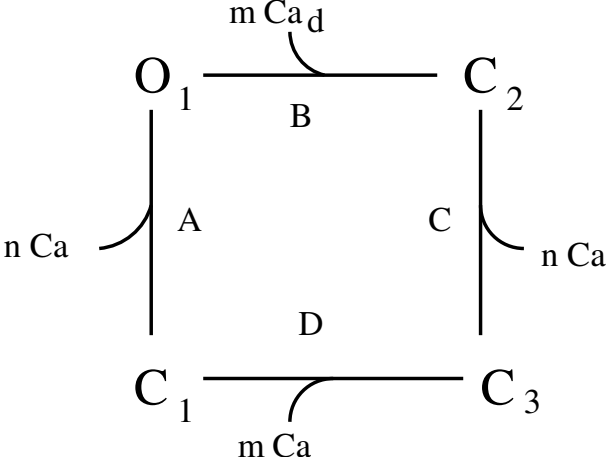


Popen



Ca

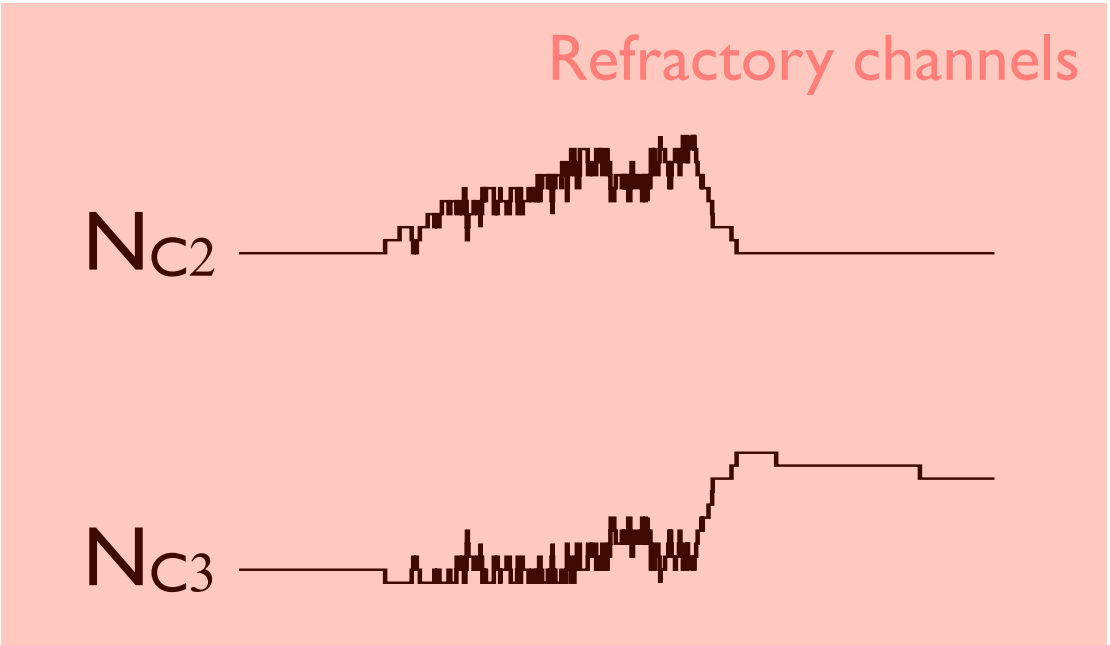
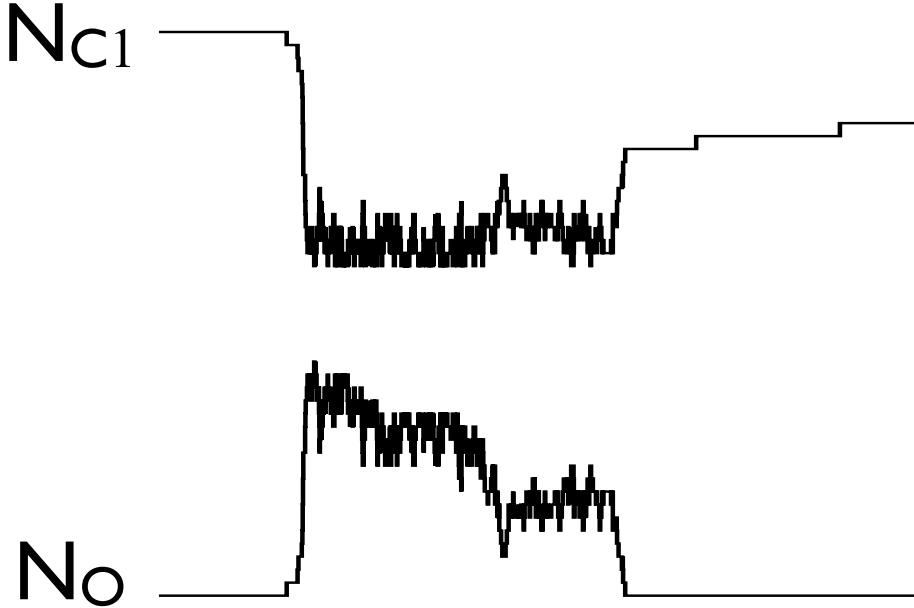
# Four-state model with $\text{Ca}^{2+}$ activation and $\text{Ca}^{2+}$ inactivation



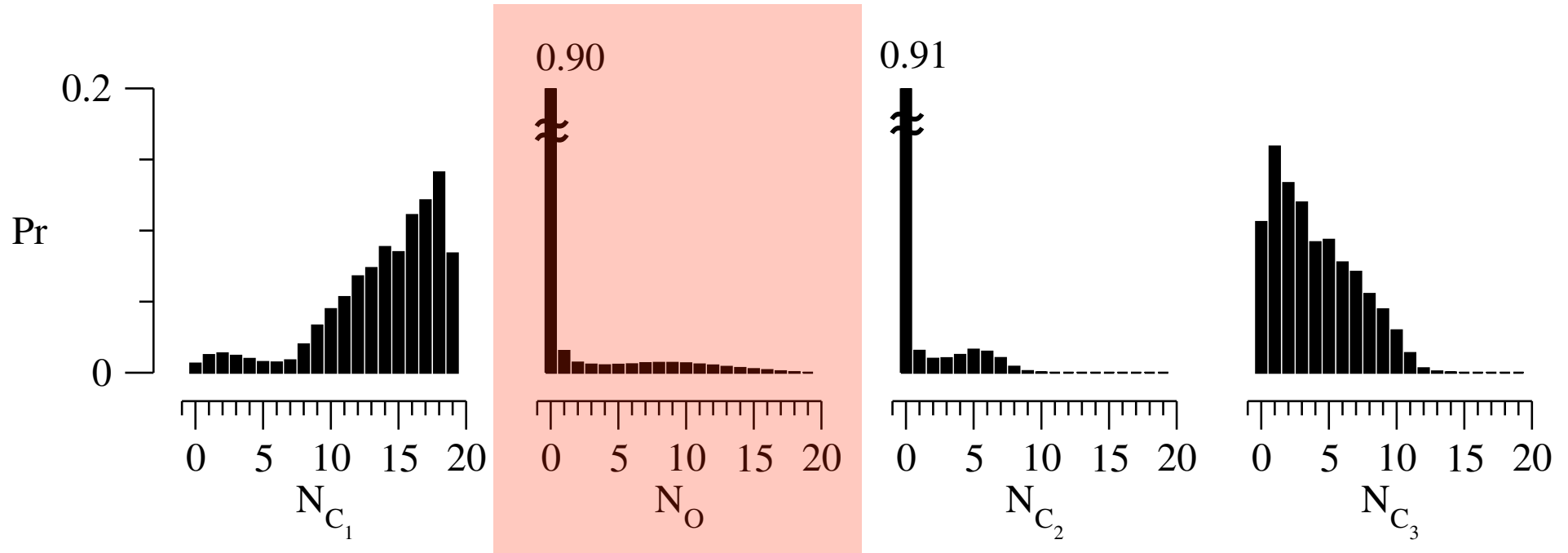
What leads to the termination of these  $\text{Ca}^{2+}$  puffs?



accumulation of Ca inactivation leads to termination of these puffs



# Limiting probability distributions



$$f_O = \frac{N_O}{N}$$

$$Score = \frac{\text{Var}[f_O]}{\text{E}[f_O]} \approx 0.5$$

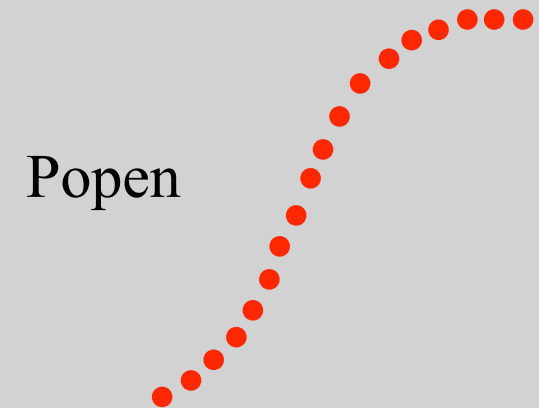
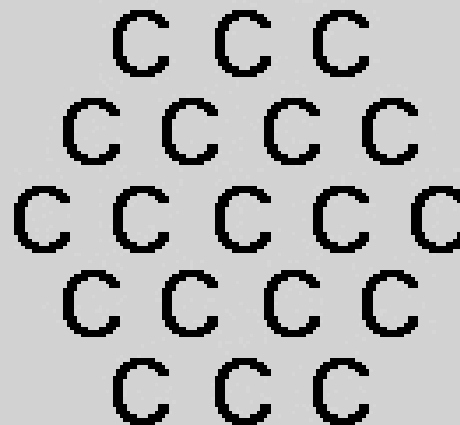
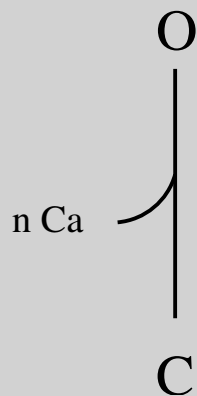
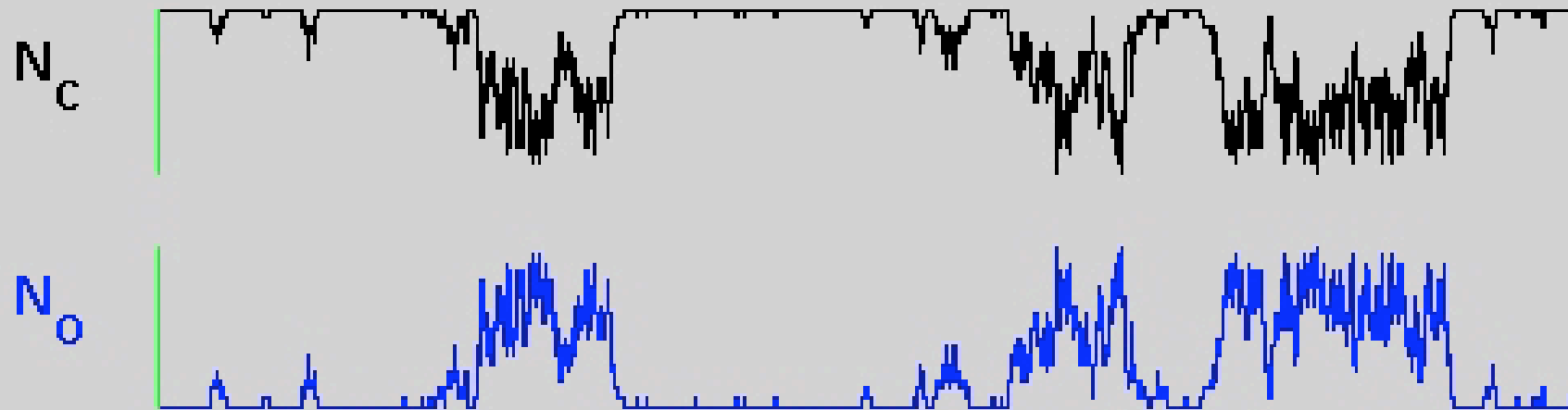
Not all parameters lead to puffs ( $Score > 0.3$ )  
but parameters leading to puffs are easy to find

# Two-state model with $\text{Ca}^{2+}$ activation but no $\text{Ca}^{2+}$ inactivation

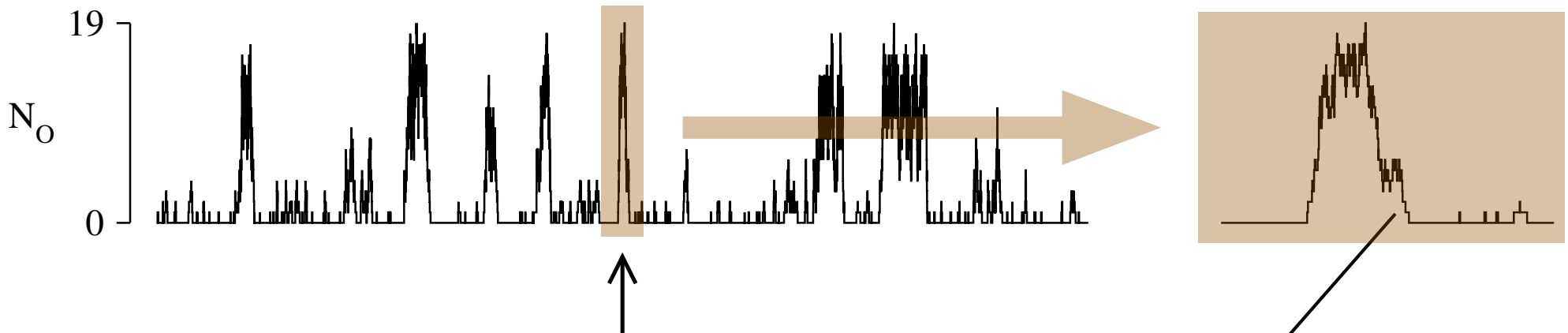
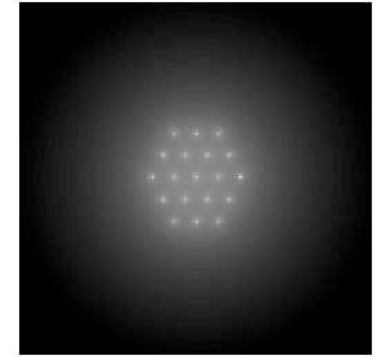
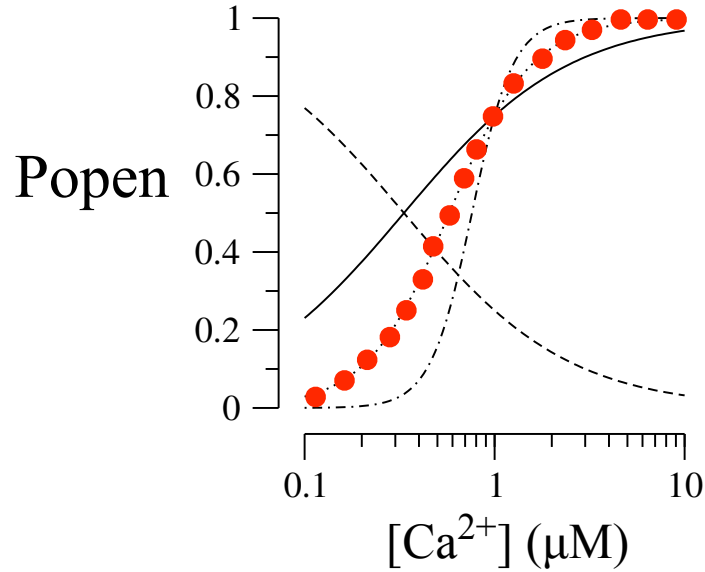
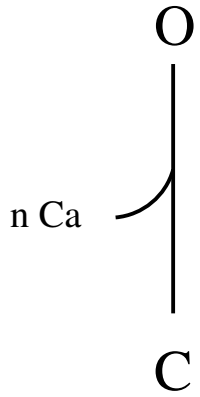
$R = 0.42 \lambda$

Score = 0.37

Time = 0 ms



# Two-state model with $\text{Ca}^{2+}$ activation but no $\text{Ca}^{2+}$ inactivation



What leads to the termination of these  $\text{Ca}^{2+}$  puffs?

Stochastic attrition??  
(cf. Cheng and Stern 2005)

# Puffs are sensitive to channel density (w/o Ca inactivation)

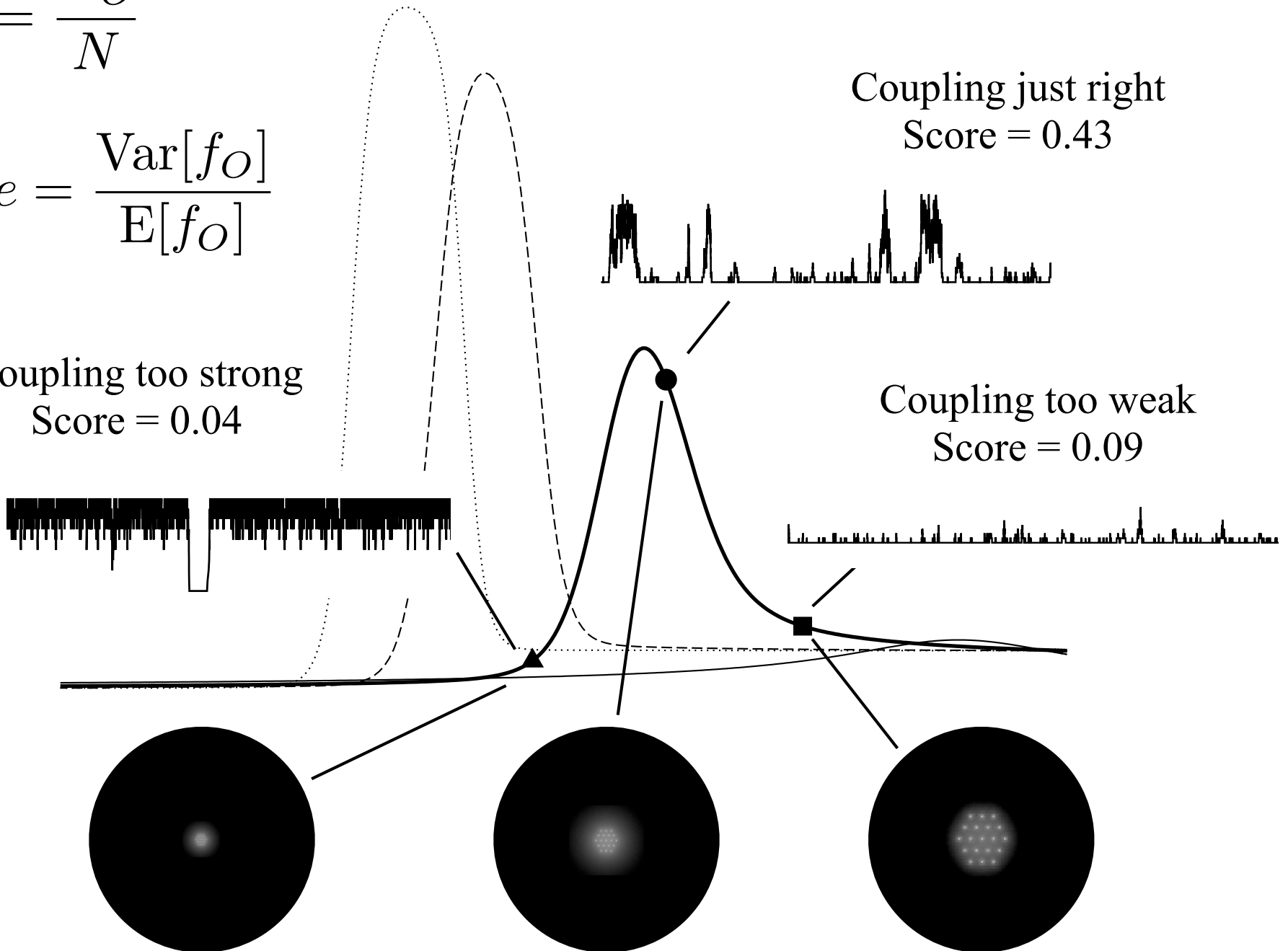
$$f_o = \frac{N_o}{N}$$

$$Score = \frac{\text{Var}[f_o]}{\text{E}[f_o]}$$

Coupling too strong  
Score = 0.04

Coupling just right  
Score = 0.43

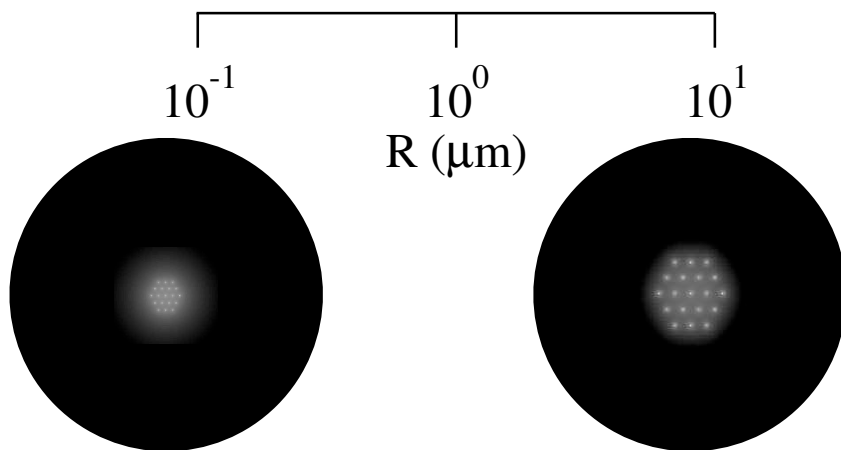
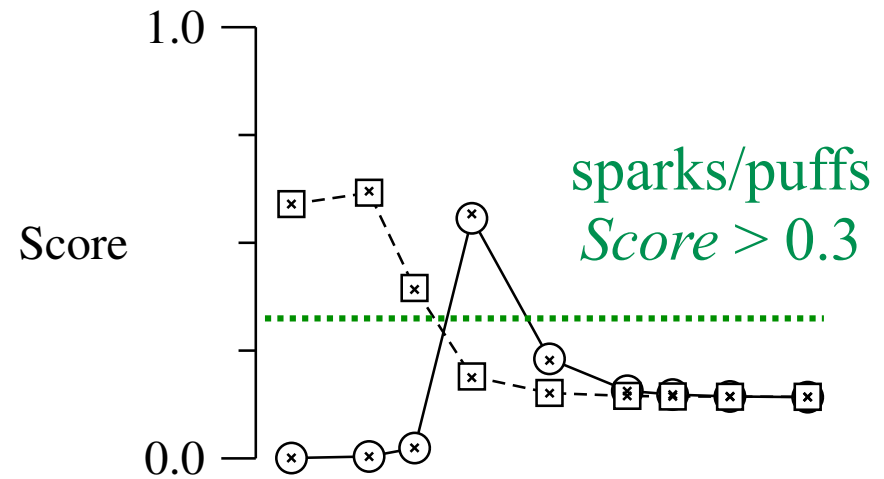
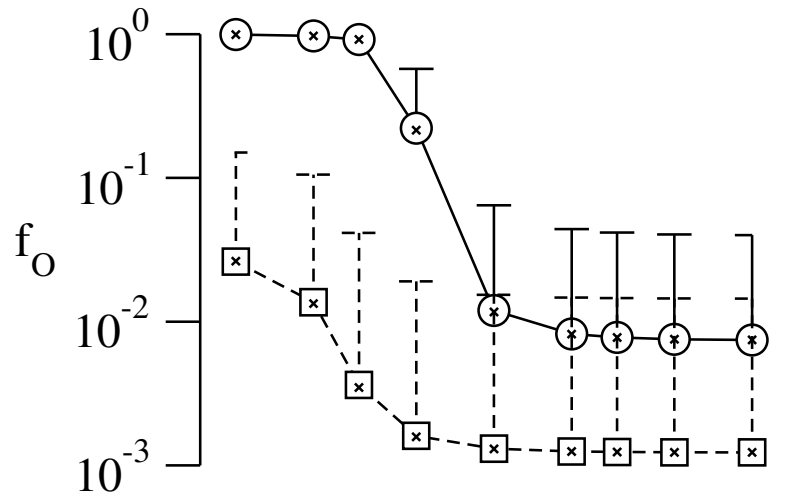
Coupling too weak  
Score = 0.09



# Puffs terminating via Ca inactivation are not sensitive to channel density

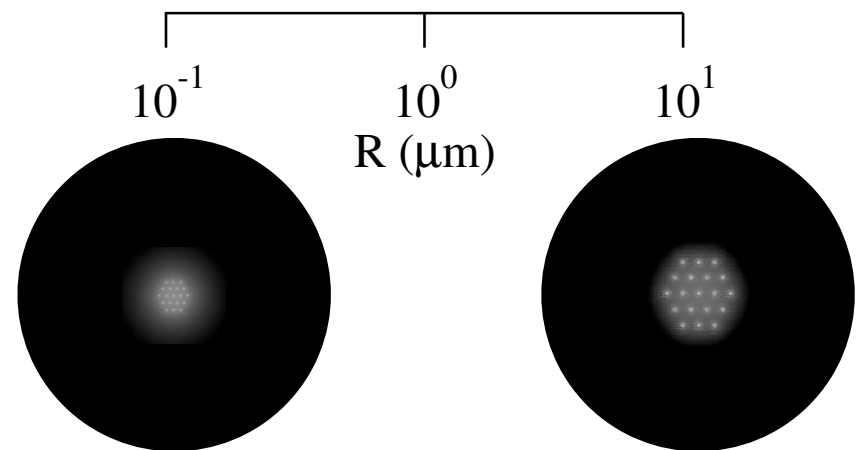
$$f_O = \frac{N_O}{N}$$

$$Score = \frac{\text{Var}[f_O]}{\text{E}[f_O]}$$



$R < \lambda$

$R > \lambda$



$R < \lambda$

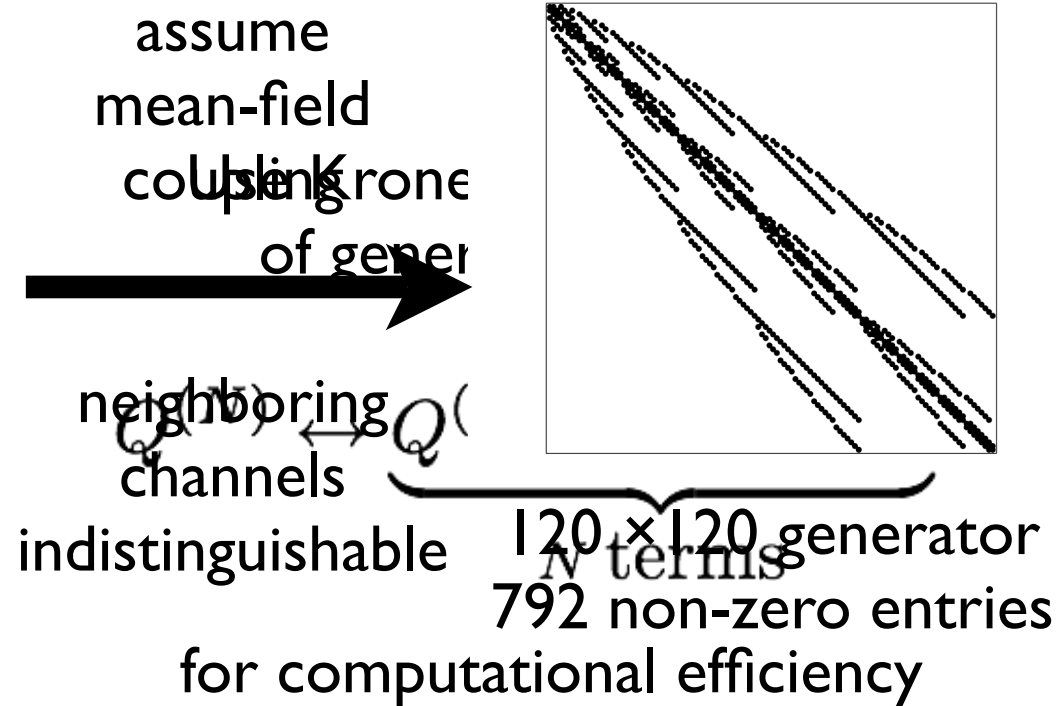
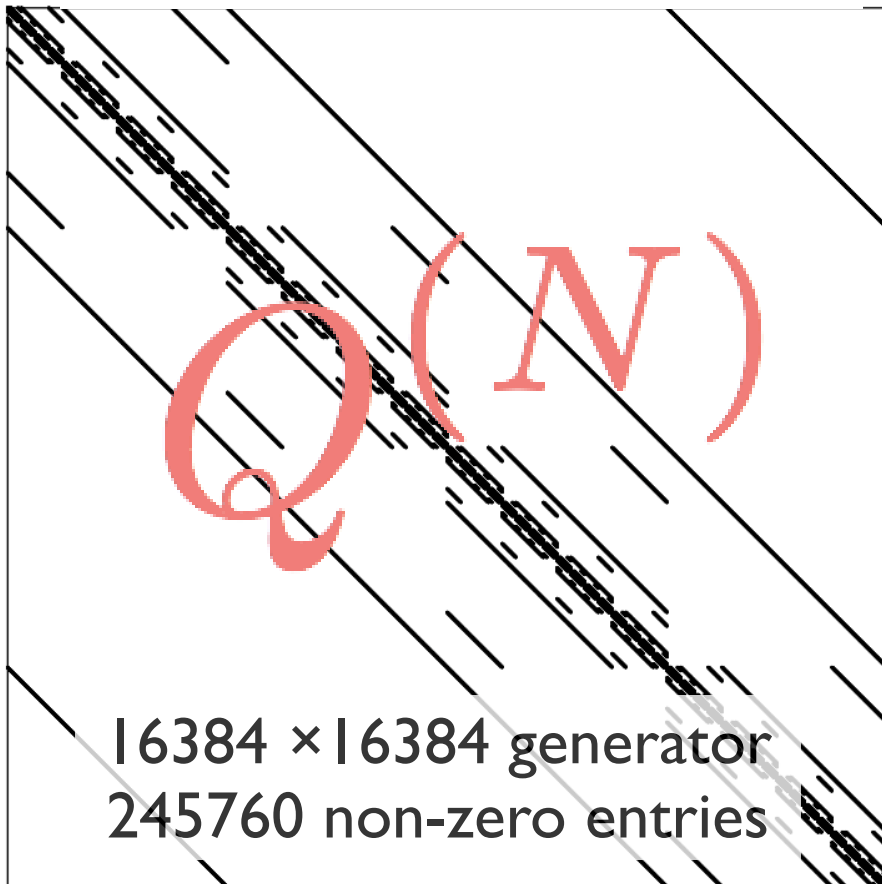
$R > \lambda$

Parameter studies are performed using direct methods

$$\pi^T Q^{(N)} = 0 \text{ subject to } \pi^T e = 1$$

- faster than Monte Carlo estimates —
- nontrivial due to state space explosion —

seven four-state channels

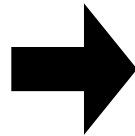


# “mean-field” approximation for channel coupling

Coupling matrix  $C = (c_{ij})$  gives  $[Ca]$  increase experienced by channel  $j$  when channel  $i$  is open

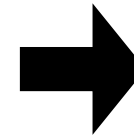
$c_{11}$	$c_{12}$	$c_{13}$	$c_{14}$
$c_{21}$	$c_{22}$	$c_{23}$	$c_{24}$
$c_{31}$	$c_{32}$	$c_{33}$	$c_{34}$
$c_{41}$	$c_{42}$	$c_{43}$	$c_{44}$

general form



$c_d$	$c_{12}$	$c_{13}$	$c_{14}$
$c_{21}$	$c_d$	$c_{23}$	$c_{24}$
$c_{31}$	$c_{32}$	$c_d$	$c_{34}$
$c_{41}$	$c_{42}$	$c_{43}$	$c_d$

channels identical



$c_d$	$c_*$	$c_*$	$c_*$
$c_*$	$c_d$	$c_*$	$c_*$
$c_*$	$c_*$	$c_d$	$c_*$
$c_*$	$c_*$	$c_*$	$c_d$

mean field

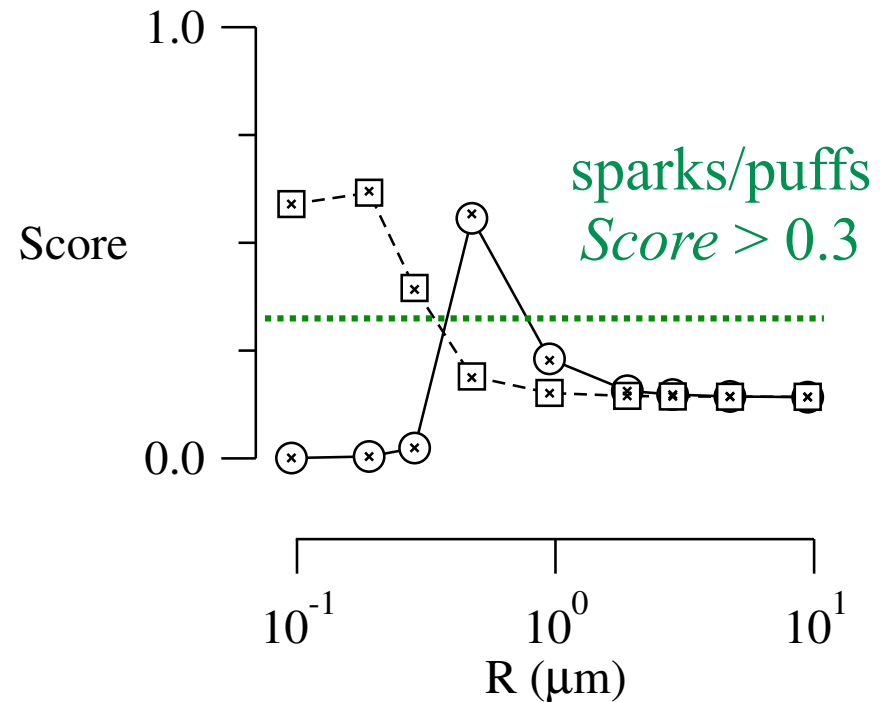
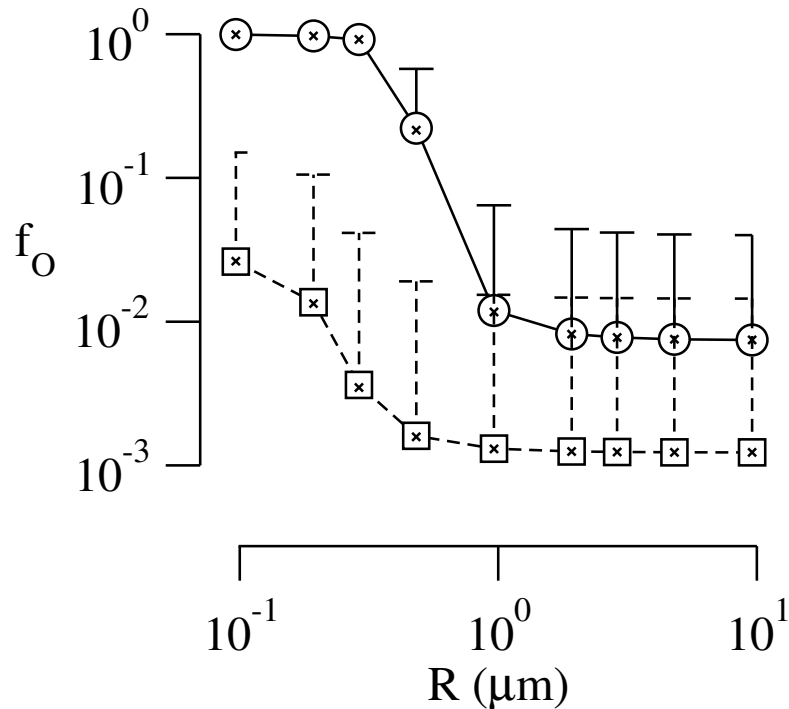
Mean-field approximation usually works well  
(see next slide)



Open symbols are full model while X's are mean-field result

$$f_O = \frac{N_O}{N}$$

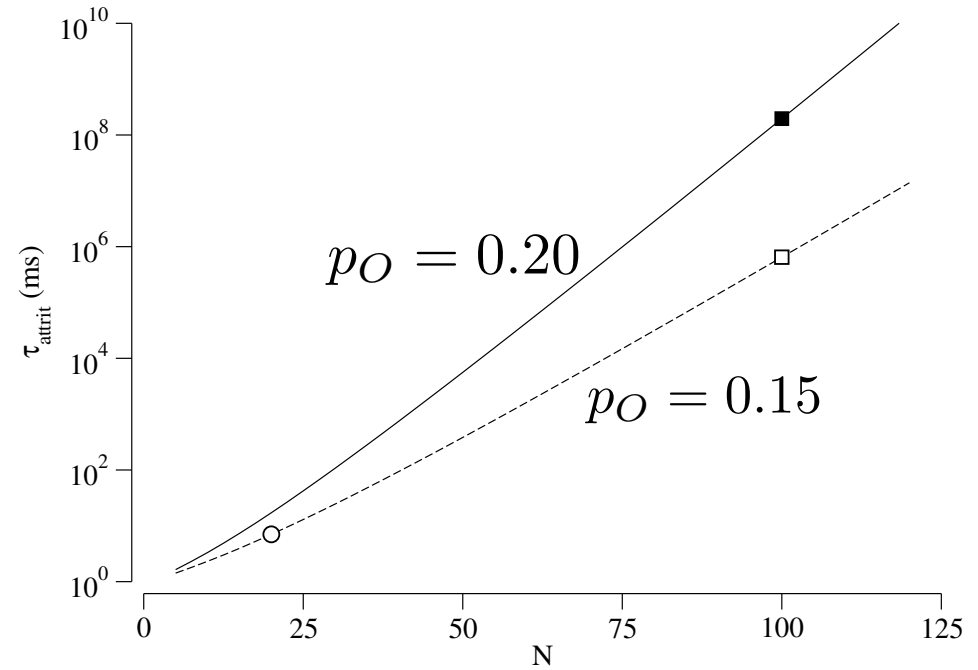
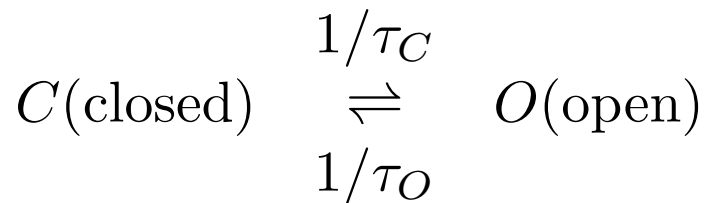
$$Score = \frac{\text{Var}[f_O]}{\text{E}[f_O]}$$



The details of channel position are important primarily through their effect on the average coupling strength ( $c^*$ )

# Isn't stochastic attrition an unlikely termination mechanism?

N two-state channels  
 identical  
 independent gating



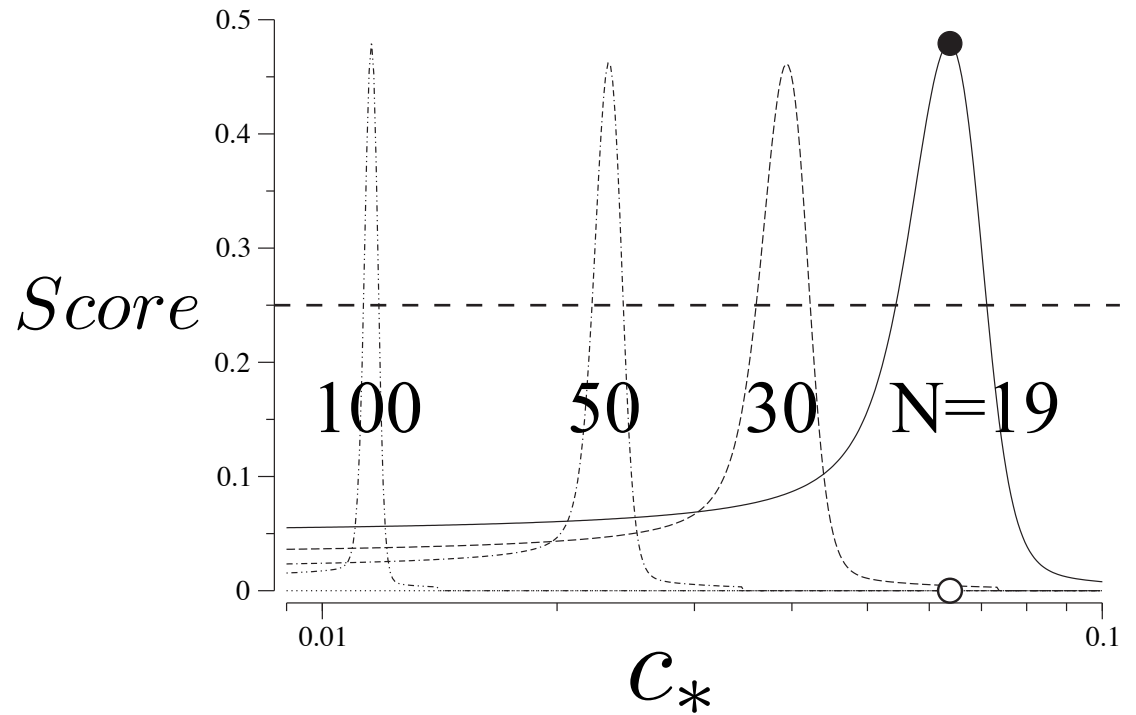
The time constant for stochastic attrition is an exponential function of the number of open channels

$$\tau_{\text{attrit}} = \frac{1}{k_{\text{attrit}}} = \tau_O \frac{1 - (1 - p_O)^N}{N (1 - p_O)^{N-1} p_O}$$

$$p_O = \frac{\tau_O}{\tau_C + \tau_O}$$

(Stern)

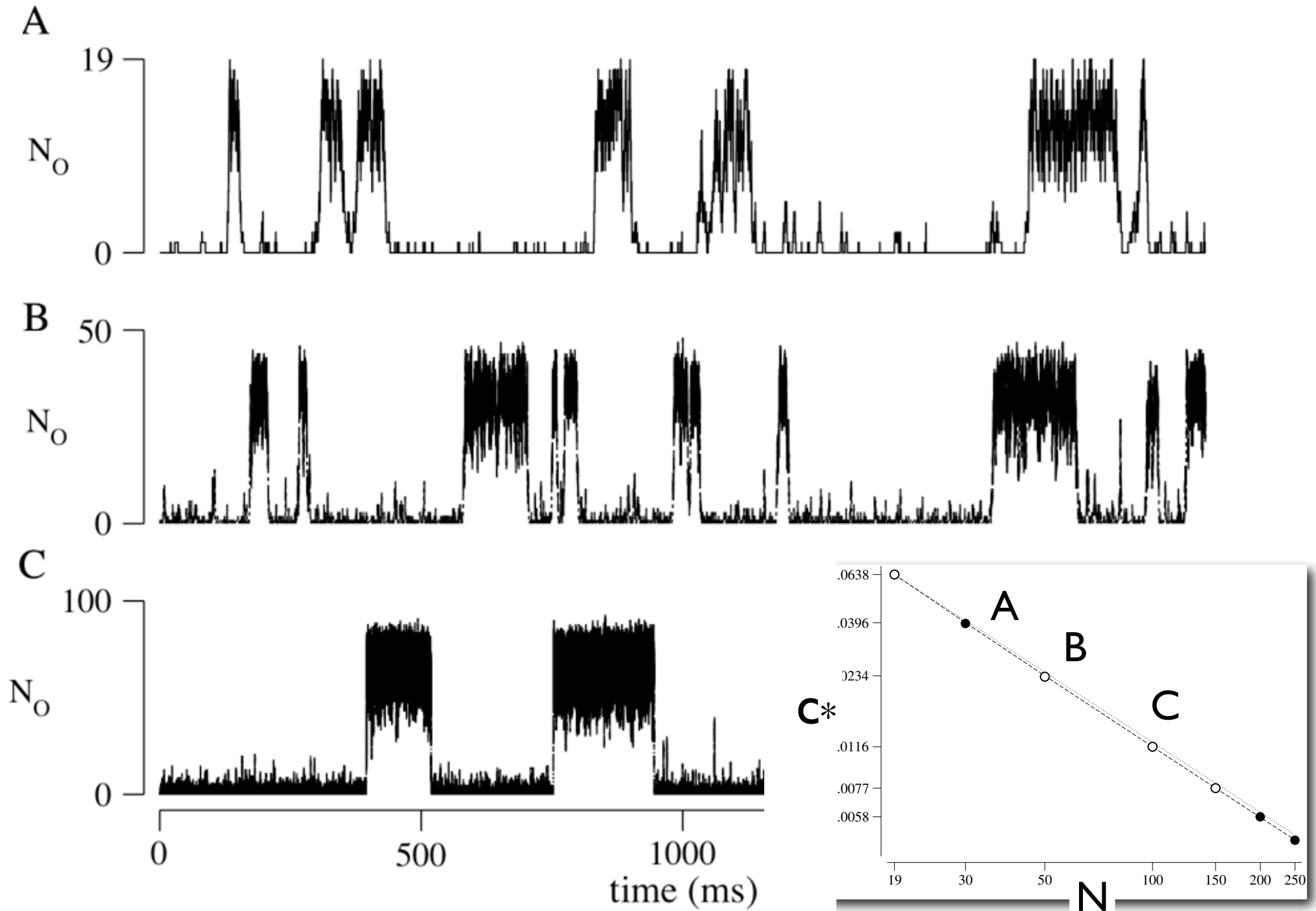
N two-state channels  
 identical  
~~independent gating~~  
 mean-field instantaneously-  
 coupled gating



The time constant for stochastic attrition depends on the coupling strength (i.e., the density of the release site)

$$\tau_{attrit} = \frac{1}{k_{attrit}} = \frac{1}{k_-} \left\{ 1 + \sum_{i=1}^{N-1} \left[ \frac{(N-1)!}{(i+1)!(N-1-i)!} \prod_{j=1}^i \left( \frac{c_\infty + jc_*}{K} \right)^\eta \right] \right\}$$

N can be large if coupling strength is appropriately reduced

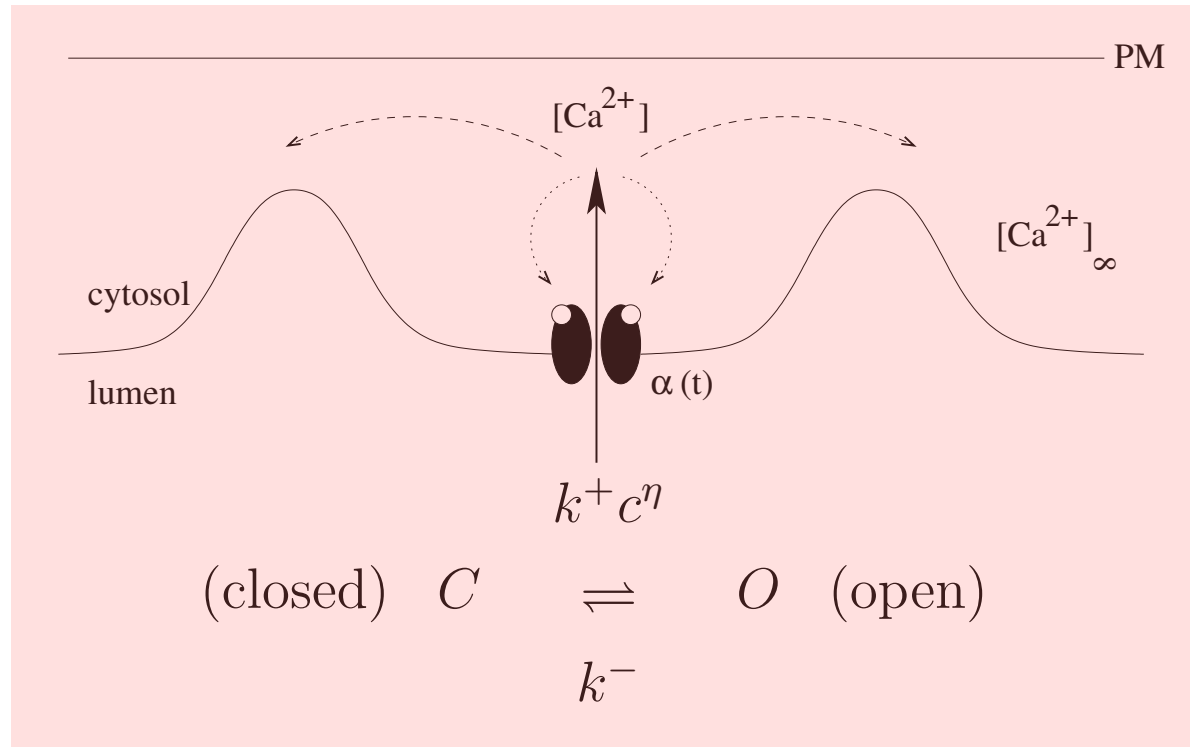


But stochastic attrition is not robust when the number of channels is large, that is, it requires channels with precisely the right density or source amplitude

One expects that the time constant for domain formation and collapse will influence puff/spark termination

Before considering Ca release sites, consider the effect of a time-dependent Ca domain on a single Ca-regulated channel

# Effect of “residual calcium” on Ca-regulated channels



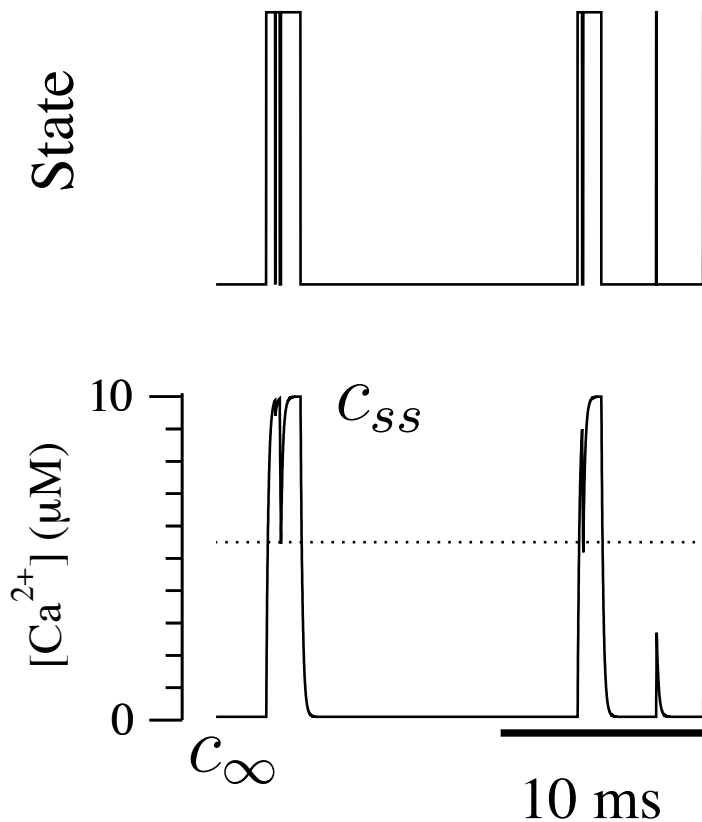
$$\frac{dc}{dt} = \alpha(t) - \frac{c - c_{\infty}}{\tau} \quad \text{where} \quad \alpha(t) = \begin{cases} 0 & \text{when } S(t) = C \\ \alpha_0 & \text{when } S(t) = O. \end{cases}$$

$$c_{\infty} < c < c_{ss} = \tau\alpha_0 + c_{\infty}$$

# Effect of “residual Ca” on Ca-activated channel

fast domain  
slow channel

$$\tau = 10^{-4}$$



increase  $\tau$



when domain is slow “residual calcium” from previous openings increases rate of  $C \rightarrow O$  transitions leading to elevated open probability

# Probability density approaches as an alternative to Monte Carlo

The joint distributions

$$\rho_C(c, t)dc = \text{P} \{ c < [\text{Ca}^{2+}] < c + dc \text{ and } S(t) = C \}$$

$$\rho_O(c, t)dc = \text{P} \{ c < [\text{Ca}^{2+}] < c + dc \text{ and } S(t) = O \}$$

are time-dependent and satisfy

$$\begin{aligned} \frac{\partial \rho_C}{\partial t} &= \left[ -\frac{\partial \phi_C}{\partial c} \right] \left[ -k^+ c^\eta \rho_C + k^- \rho_O \right] \\ \frac{\partial \rho_O}{\partial t} &= \left[ -\frac{\partial \phi_O}{\partial c} \right] \left[ +k^+ c^\eta \rho_C - k^- \rho_O \right] \end{aligned}$$

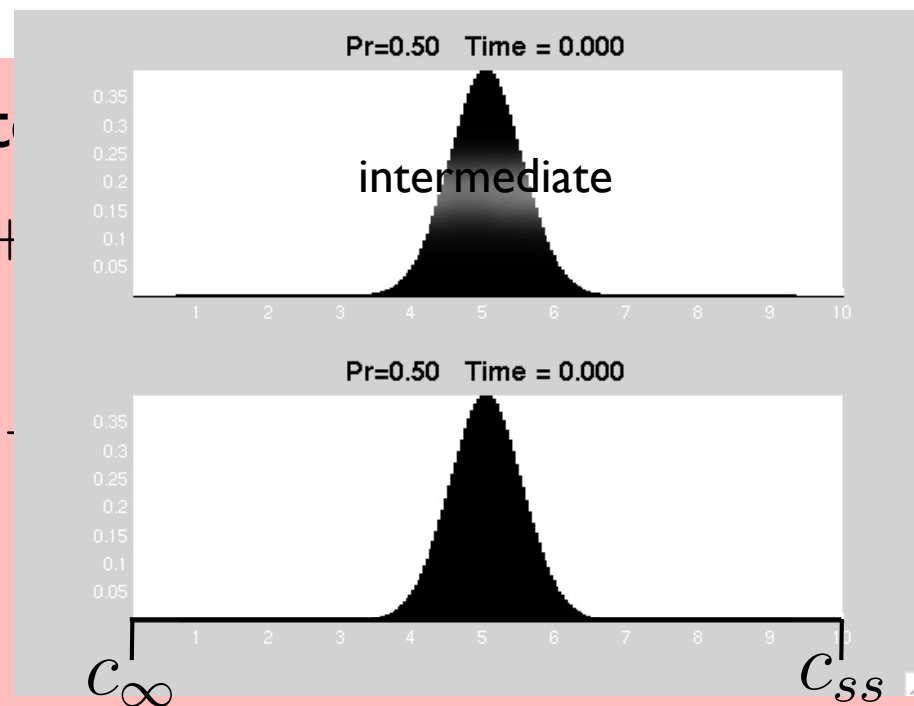
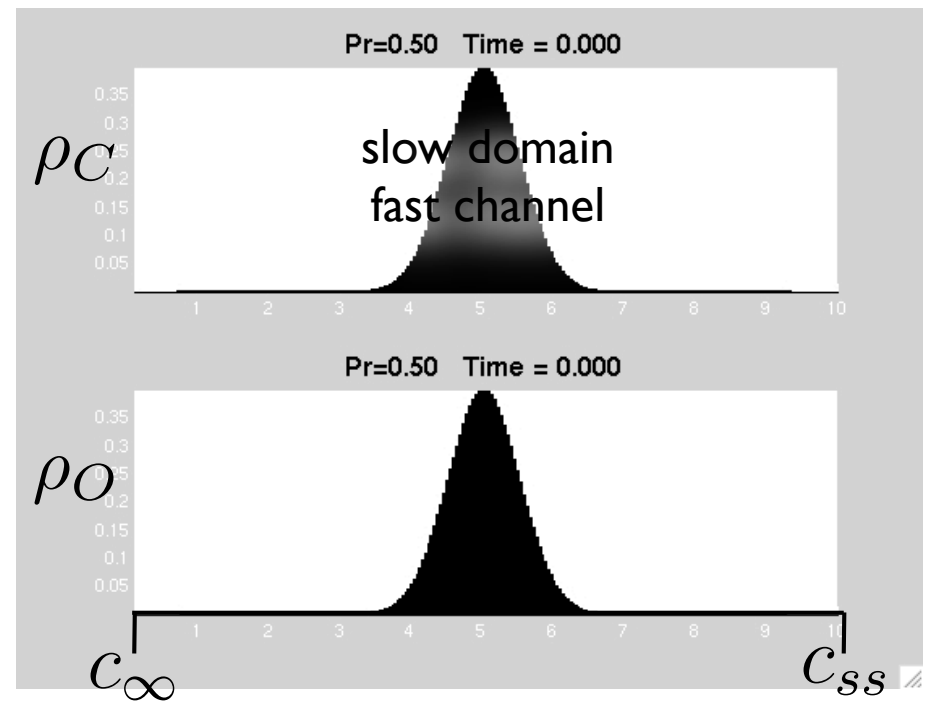
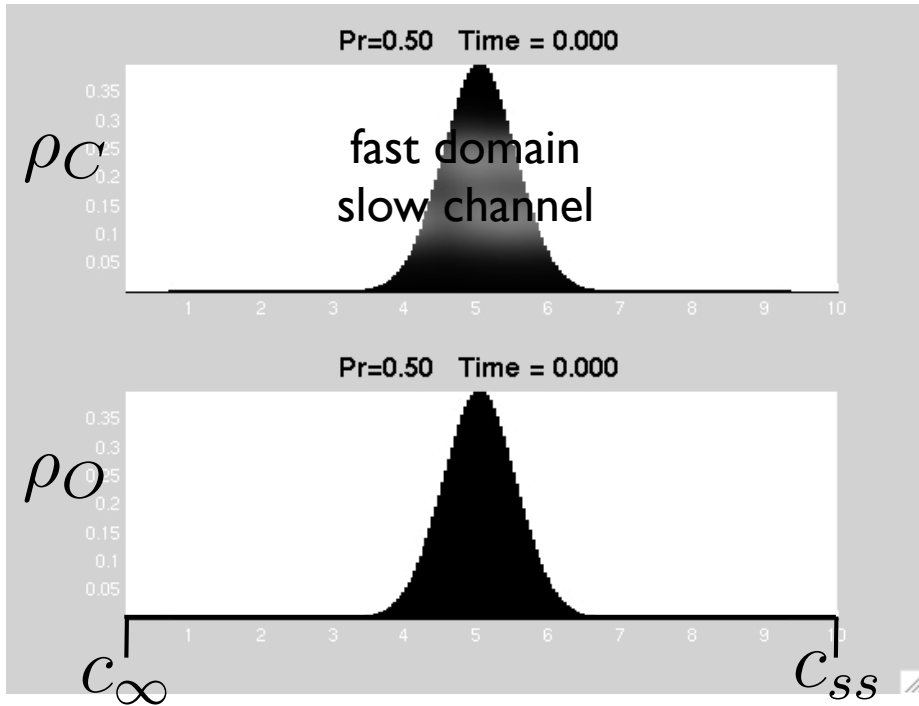
reaction (stochastic gating)

advection (deterministic dynamics of domain)

$$\begin{aligned} \phi_C(c, t) &= j_C(c) \rho_C(c, t) & j_C(c) &= -\frac{c - c_\infty}{\tau} \\ \phi_O(c, t) &= j_O(c) \rho_O(c, t) & j_O(c) &= \alpha_0 - \frac{c - c_\infty}{\tau} \end{aligned}$$



# Time-dependent probability densities



$$\rho_C = \hat{\rho} e^{\tau^+ c}$$

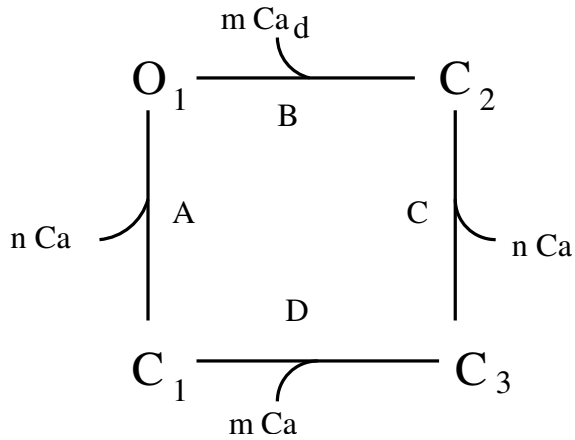
$$\rho_O = \hat{\rho} e^{\tau^- c}$$

$$) \tau^+ c_\infty - 1$$

$$c_\infty) \tau^+ c_\infty$$

$$\tau^\pm = \tau k^\pm$$

# The PD approach w/ more complicated single channel models

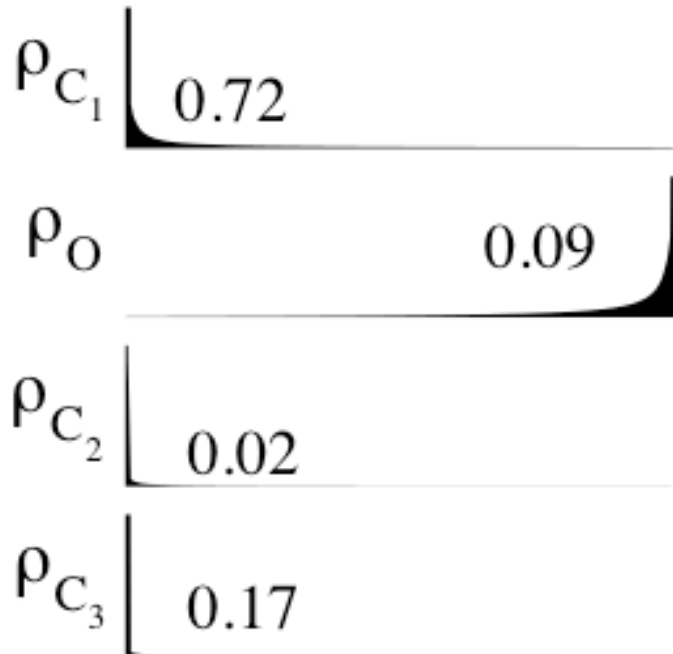


$$\rho_i(c, t)dc = P \{c < [\text{Ca}^{2+}] < c + dc \text{ and } S(t) = \mathcal{S}_i\}$$

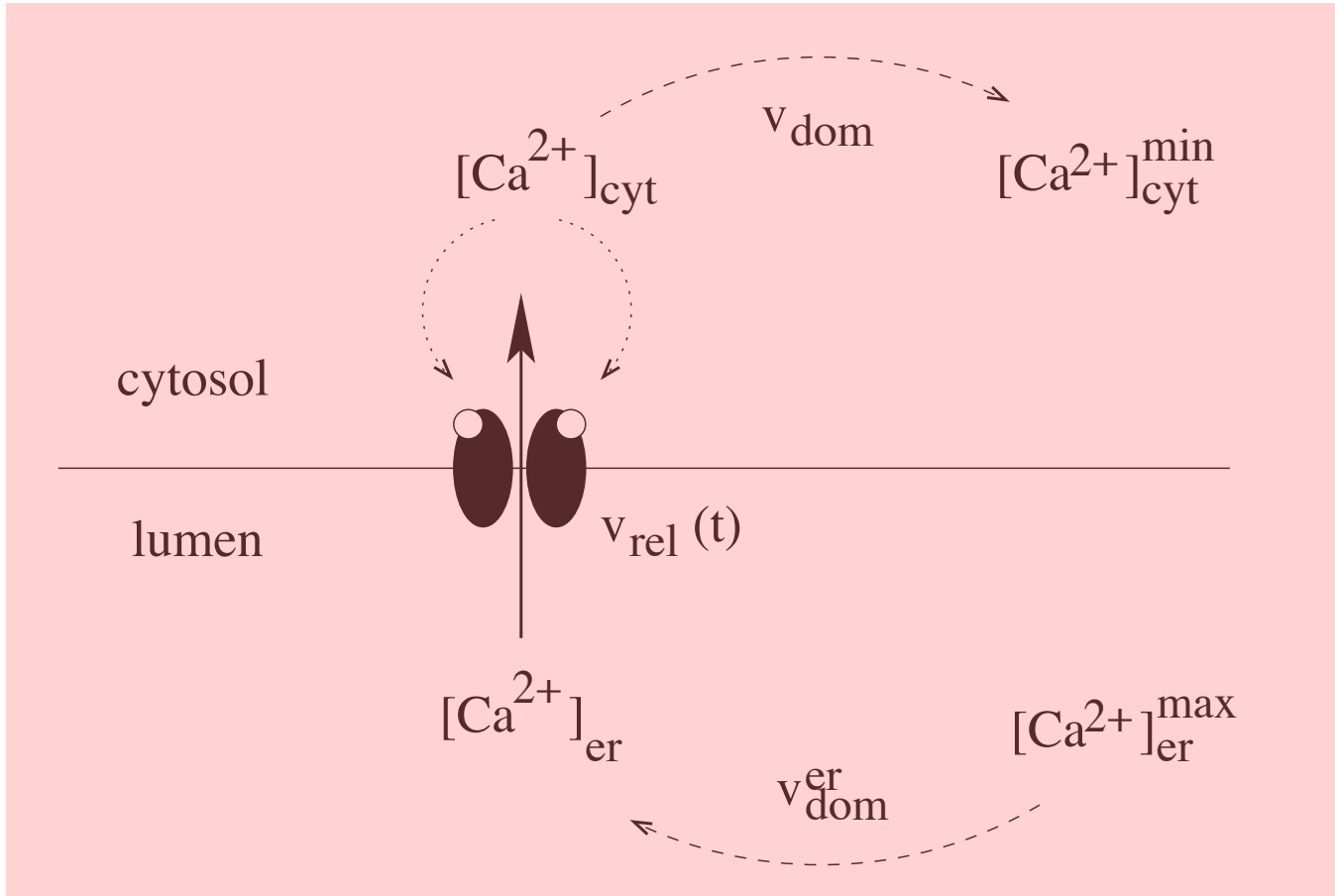
$$\frac{\partial \rho}{\partial t} = -\frac{\partial}{\partial c} [\rho J] + \rho Q$$

$$\rho = (\rho_1, \rho_2, \dots, \rho_M)$$

$\tau$  small



# The PD approach to study the effect of “luminal depletion”



$$\frac{dC_{cyt}}{dt} = \gamma(t)v_{rel}(C_{er} - C_{cyt}) - v_{dom}(C_{cyt} - C_{min})$$

$$\frac{dC_{er}}{dt} = \frac{1}{\lambda} [-\gamma(t)v_{rel}(C_{er} - C_{cyt}) + v_{dom}^{er}(C_{max} - C_{er})]$$

The joint probability densities become multivariate

$$\rho_i(C_{cyt}, C_{er}, t) dC_{cyt} dC_{er} = \text{Prob}\{C_{cyt} < \tilde{C}_{cyt} < C_{cyt} + dC_{cyt} \text{ AND} \\ C_{er} < \tilde{C}_{er} < C_{er} + dC_{er} \text{ AND } \gamma(t) = i\}$$

but still satisfy a system of **advection-reaction** equations

$$\frac{\partial}{\partial t} \rho_i = \boxed{\frac{\partial(F_{cyt}^i \rho_i)}{\partial C_{cyt}} - \frac{\partial(F_{er}^i \rho_i)}{\partial C_{er}}} + \boxed{[\vec{\rho} \mathbf{Q}]_i}$$

The probability flux has two components

$$F_{cyt}^i = \gamma_i \nu_{rel}(C_{er} - C_{cyt}) - \nu_{dom}(C_{cyt} - C_{min}) \\ F_{er}^i = \frac{1}{\lambda} [-\gamma_i \nu_{rel}(C_{er} - C_{cyt}) + \nu_{dom}^{er}(C_{max} - C_{er})]$$

each of which depends on channel state

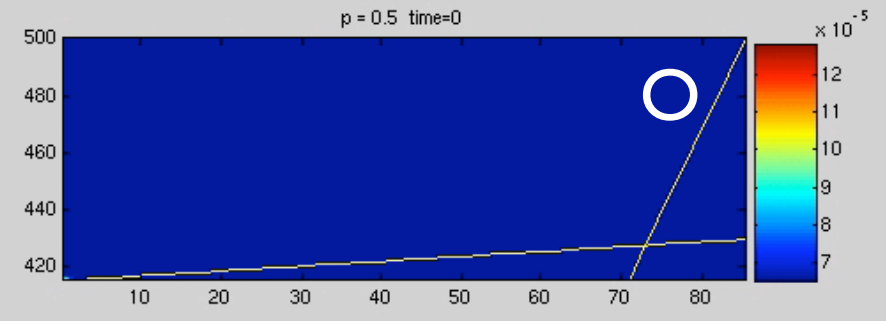
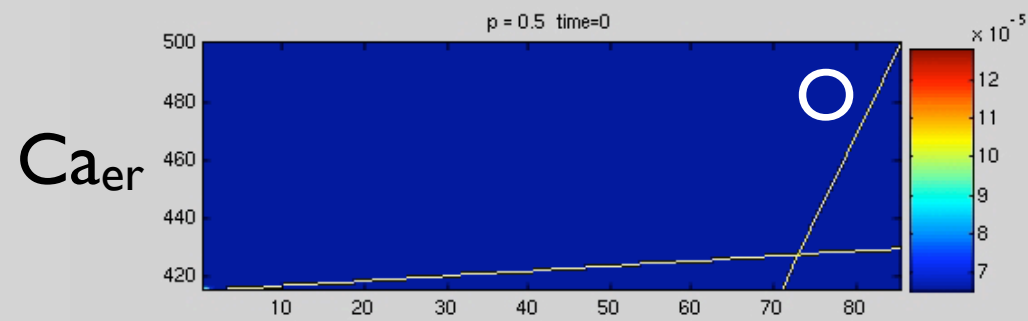
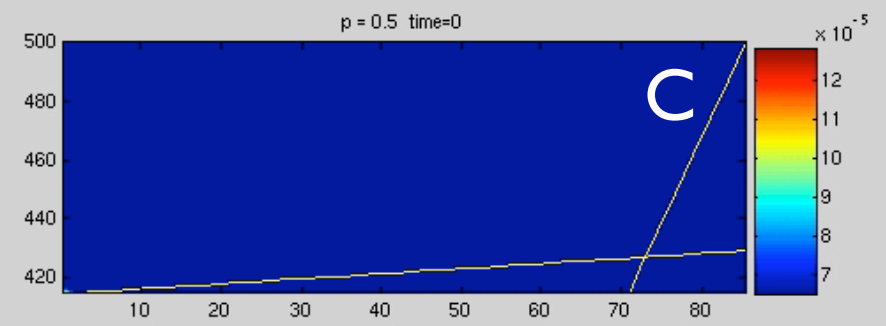
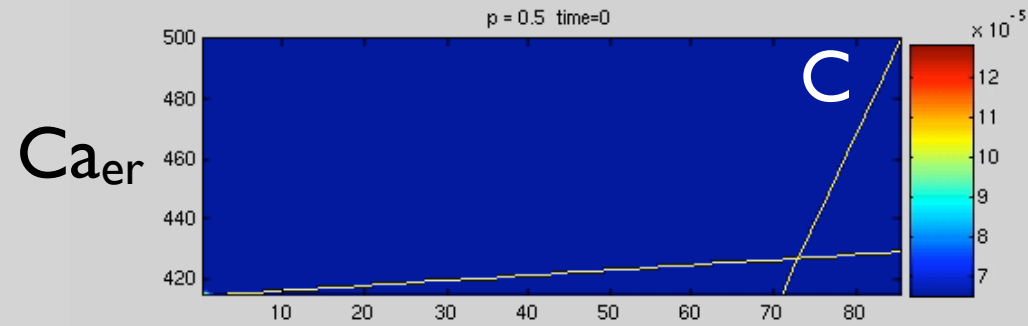
# Cytosolic domain and luminal depletion domain Ca-activated channel

fast domain  
(large  $v$ 's)

slow channel

slow domain  
(small  $v$ 's)

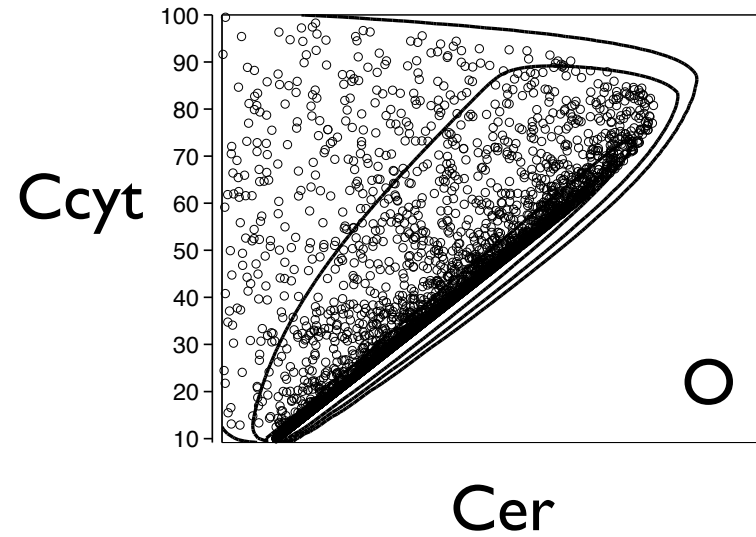
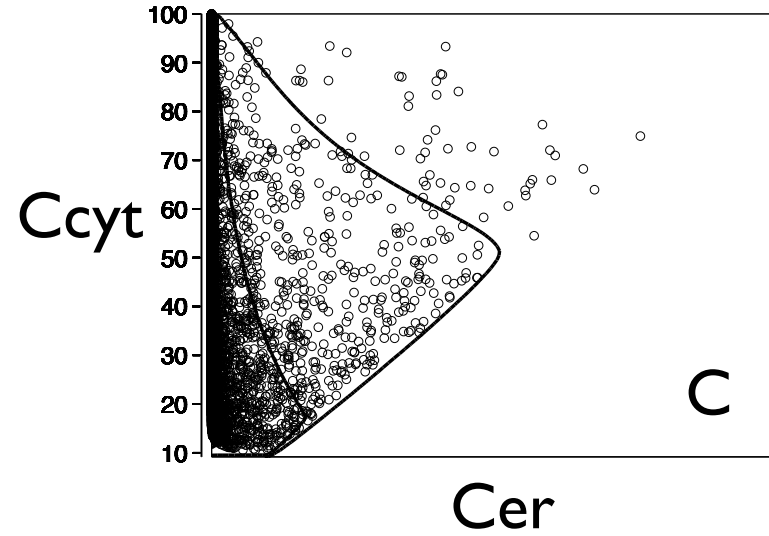
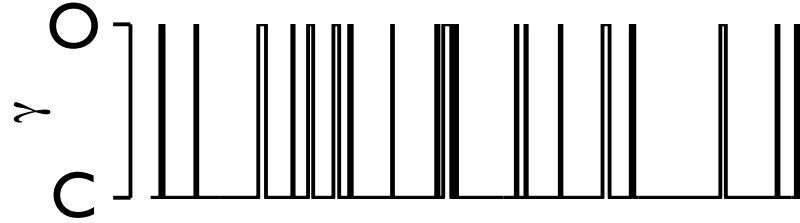
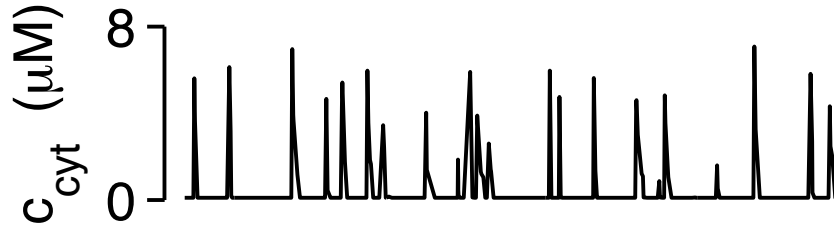
fast channel



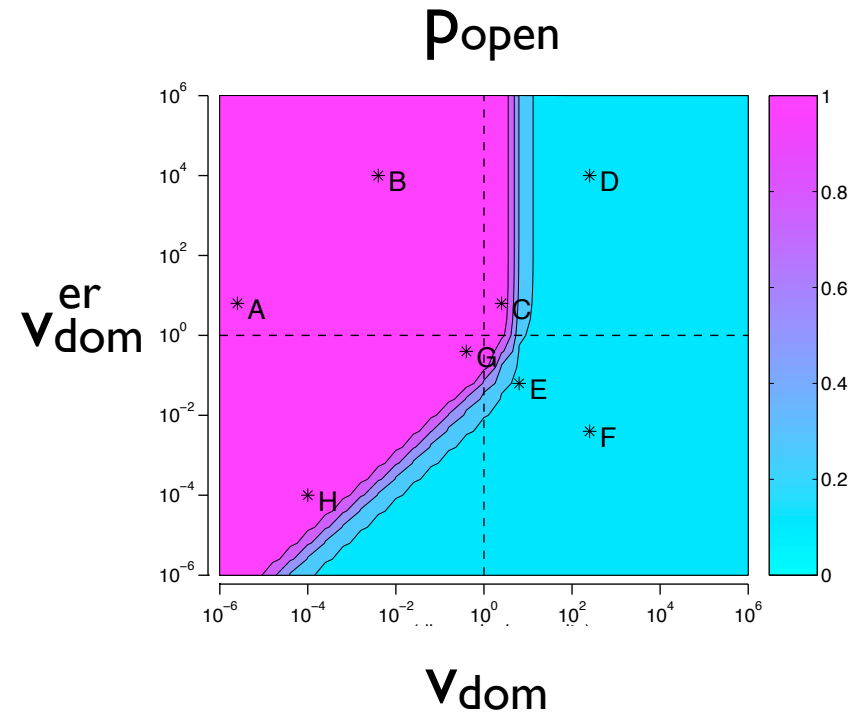
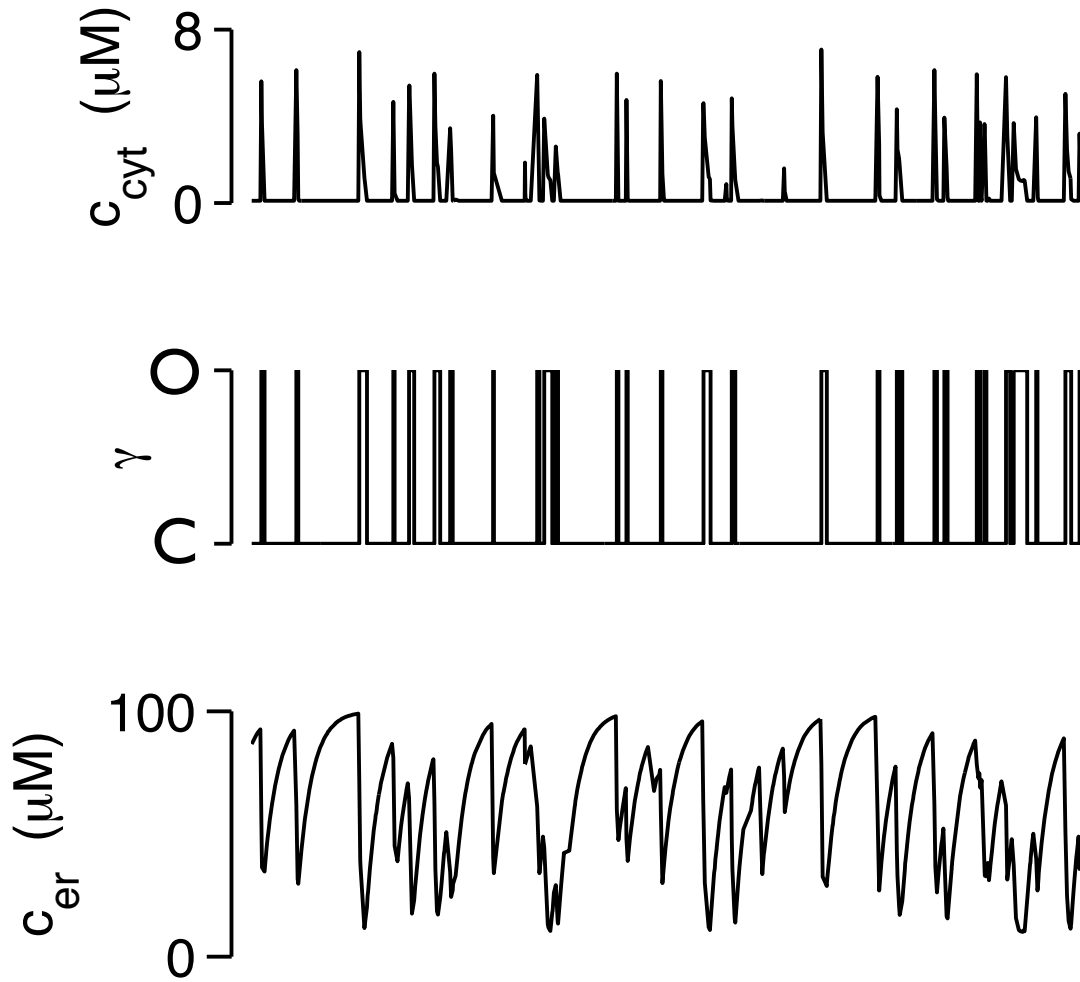
$Ca_{cyt}$

$Ca_{cyt}$

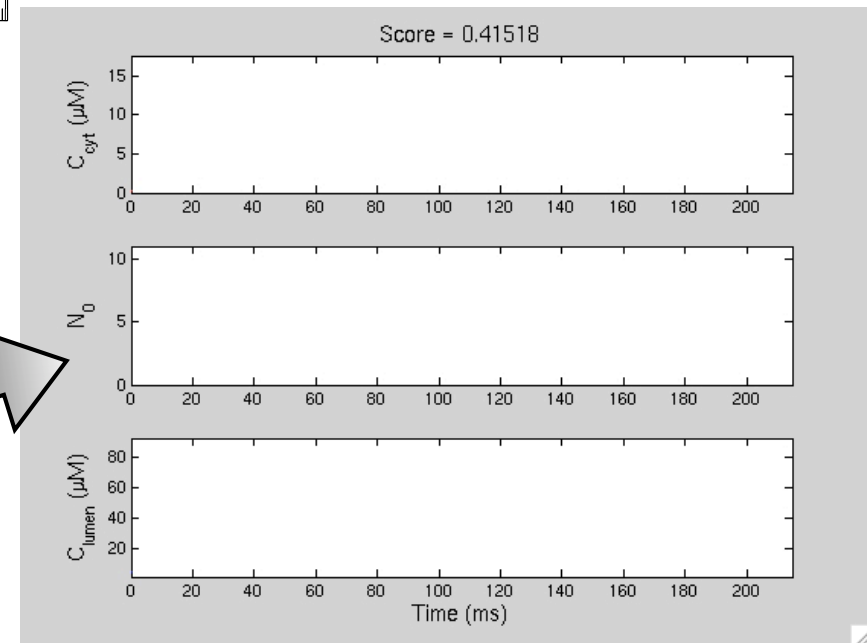
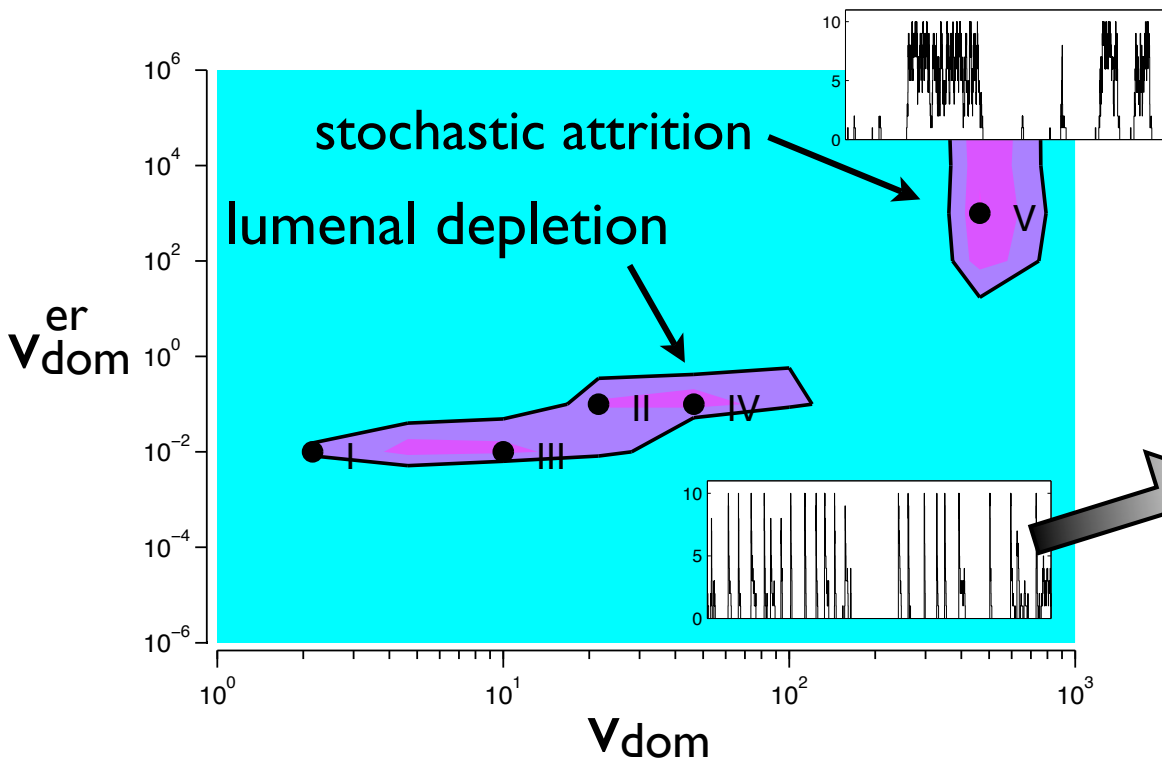
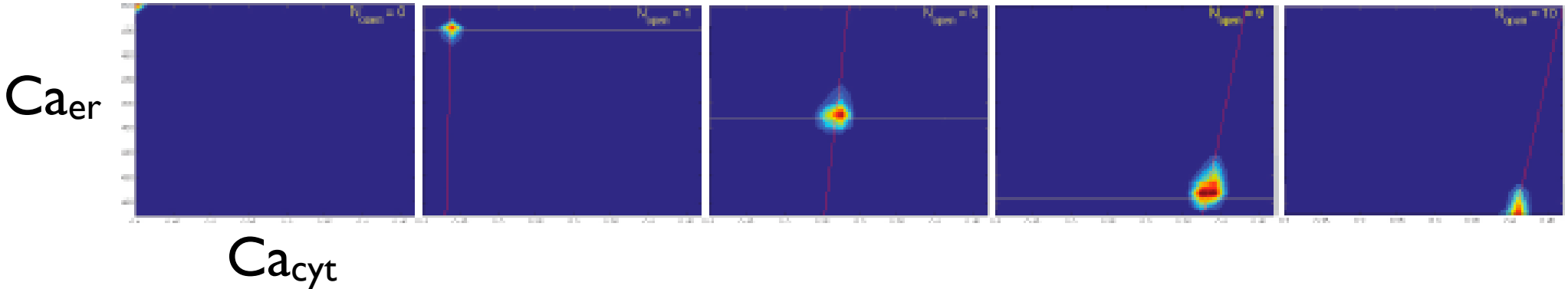
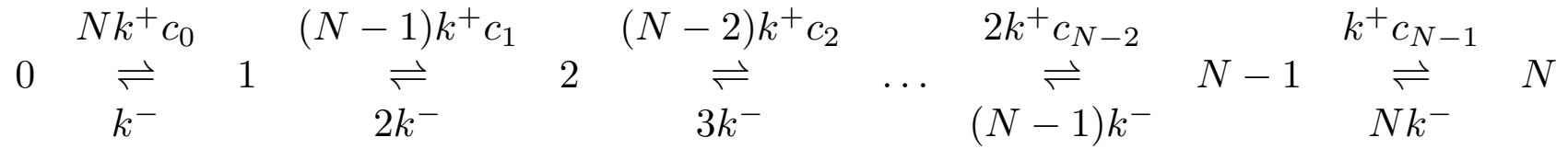
# Comparison of Monte Carlo and probability density approaches



# Calcium activated channel — effect of luminal depletion



# Dynamics of spark termination — effect of luminal depletion

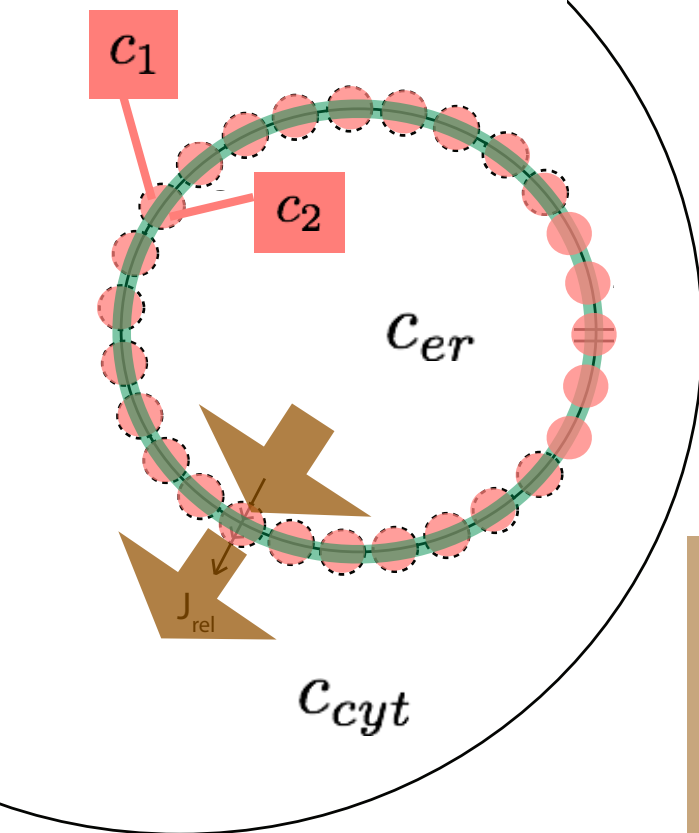




# A new class of whole cell models

not explicitly spatial  
but includes local signaling

$$\frac{\partial \rho}{\partial t} = - \frac{\partial}{\partial c} [\rho J] + \rho Q$$



$$\frac{dc_{cyt}}{dt} = J_{cyt}^* + J_{leak} - J_{pump}$$

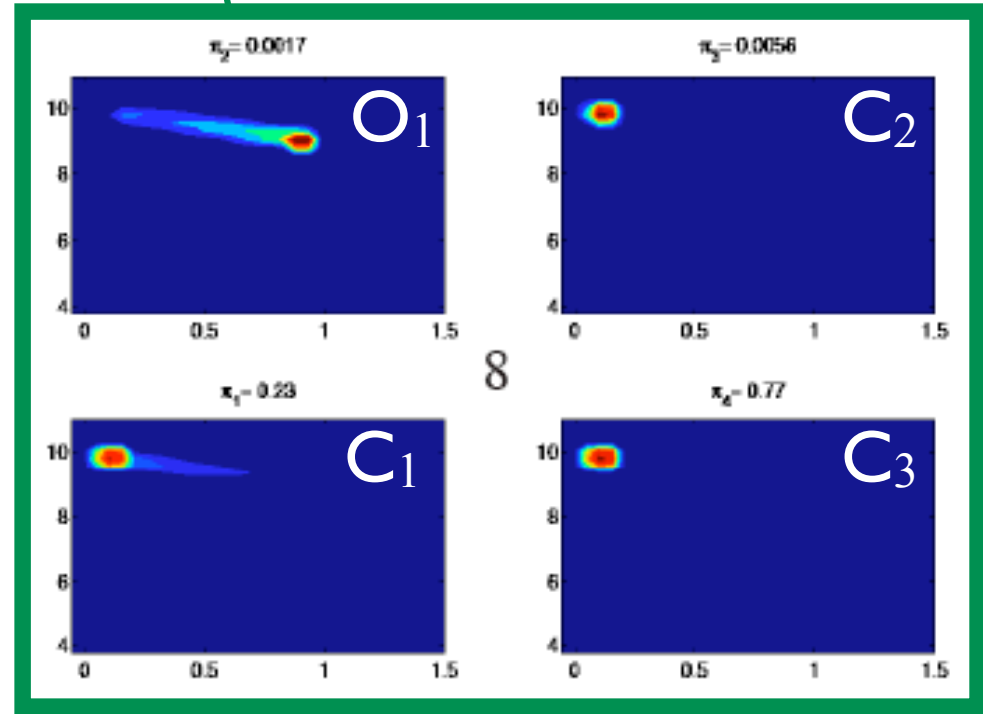
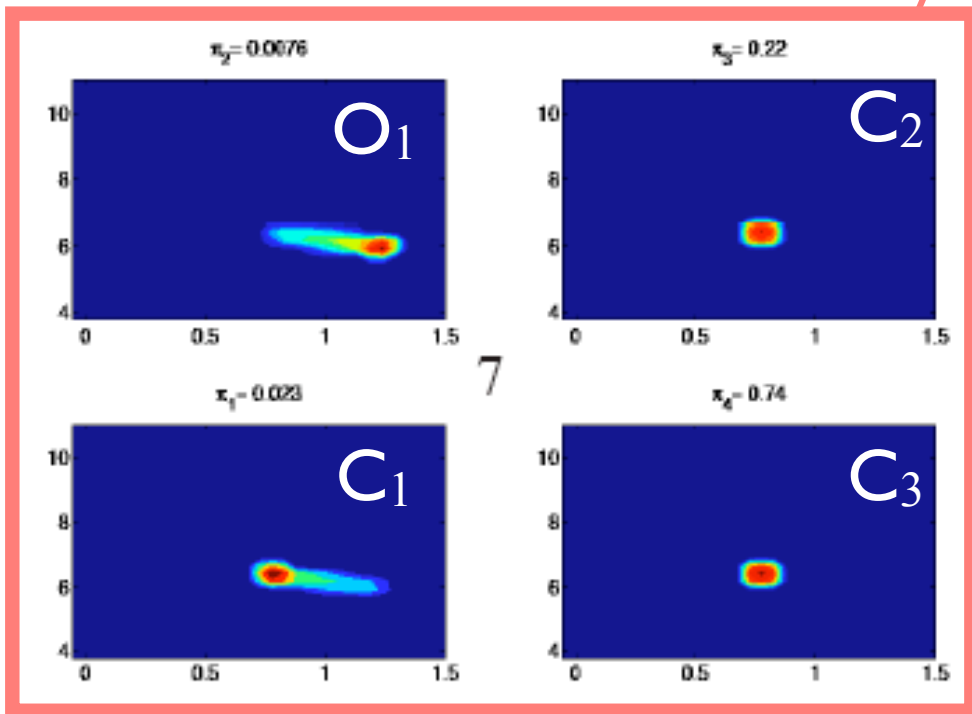
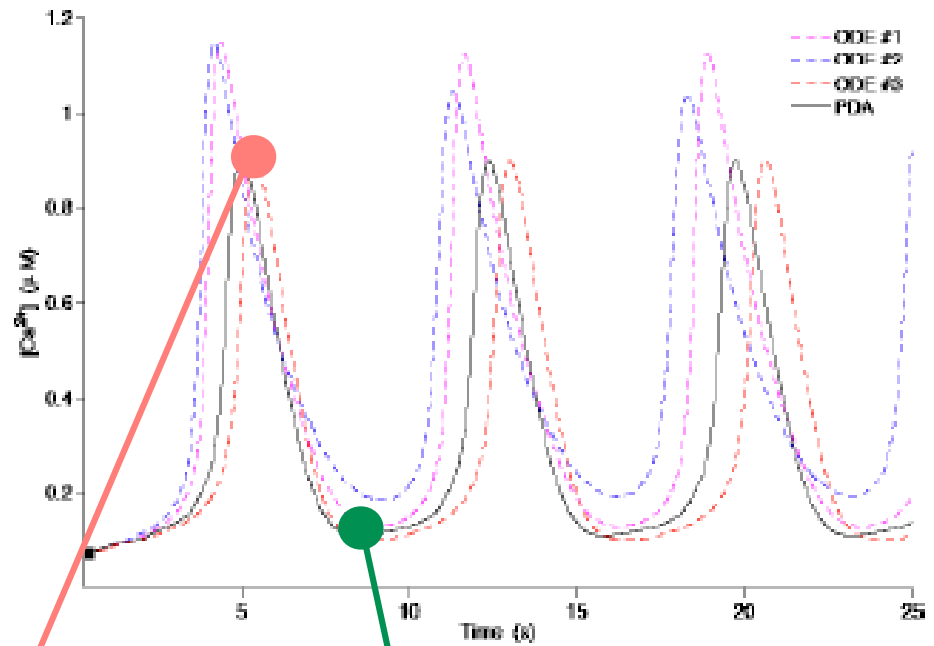
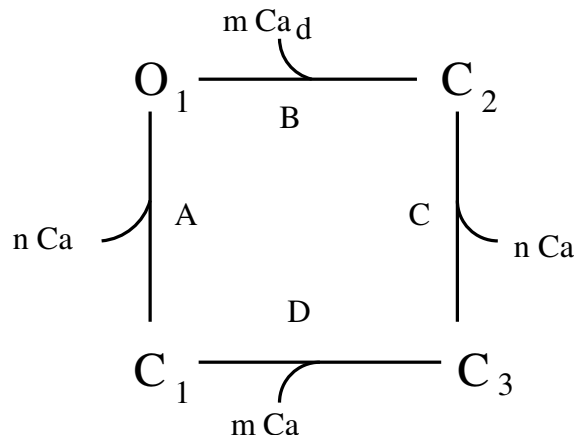
$$\frac{dc_{er}}{dt} = \frac{1}{\lambda_{er}} (J_{pump} - J_{leak} - J_{er}^*)$$

$$J_{cyt}^* = \sum_{i=1}^M \int_{c_2^{min}}^{c_2^{max}} \int_{c_1^{min}}^{c_1^{max}} v_{cyt}(c_1 - c_{cyt}) \rho_i(c_1, c_2) dc_1 dc_2$$

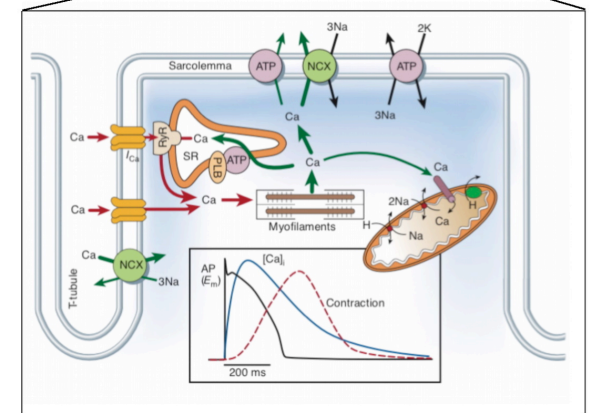
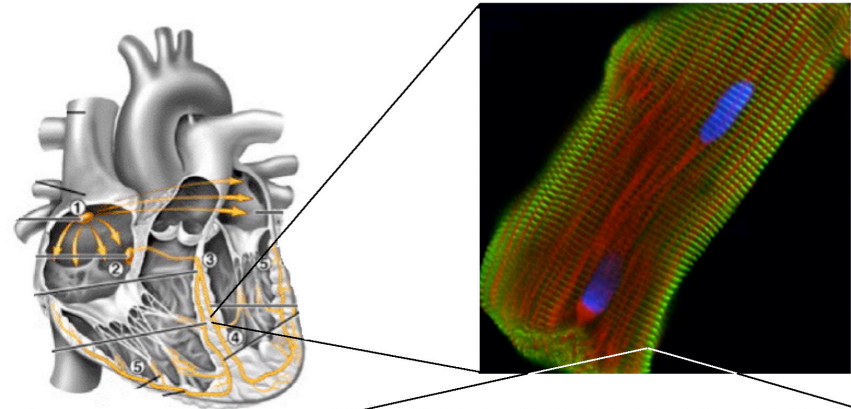
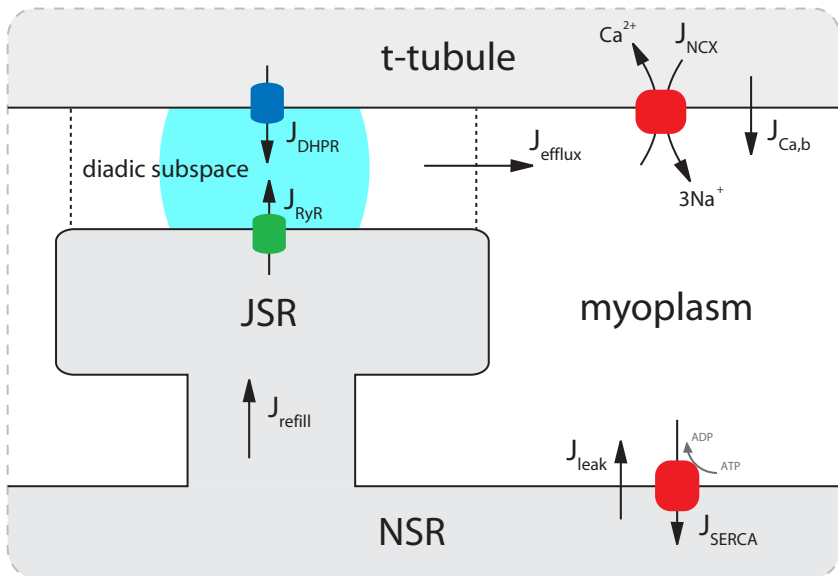
$$J_{er}^* = \sum_{i=1}^M \int_{c_2^{min}}^{c_2^{max}} \int_{c_1^{min}}^{c_1^{max}} v_{er}(c_{er} - c_2) \rho_i(c_1, c_2) dc_1 dc_2$$

large number of channels  
each with it's own time-dependent domain

# Diffuse IP3Rs with time-dependent domains

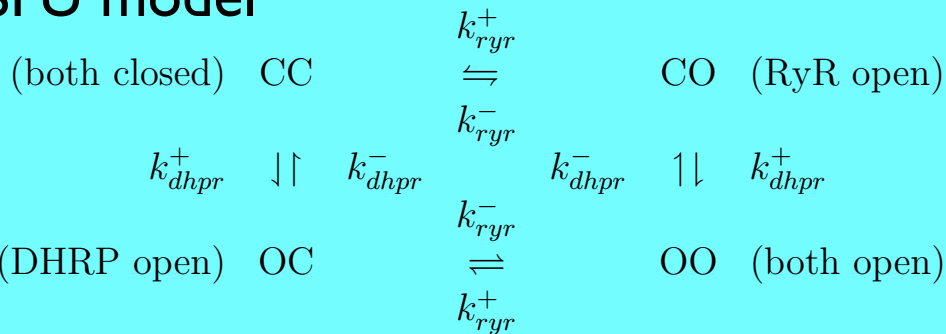


# New approach for modeling “local control” during EC coupling



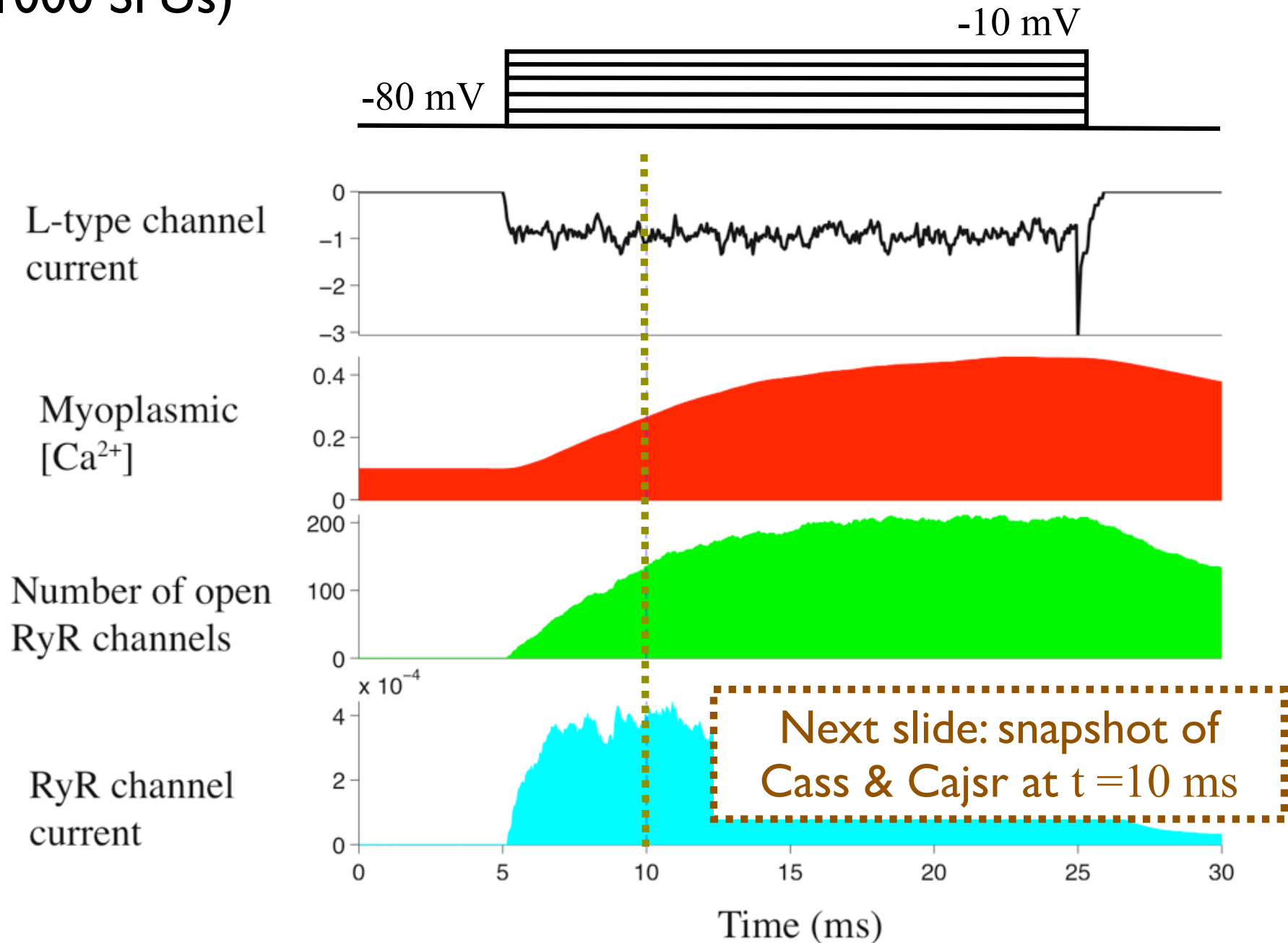
Bers 2002

## SFU model

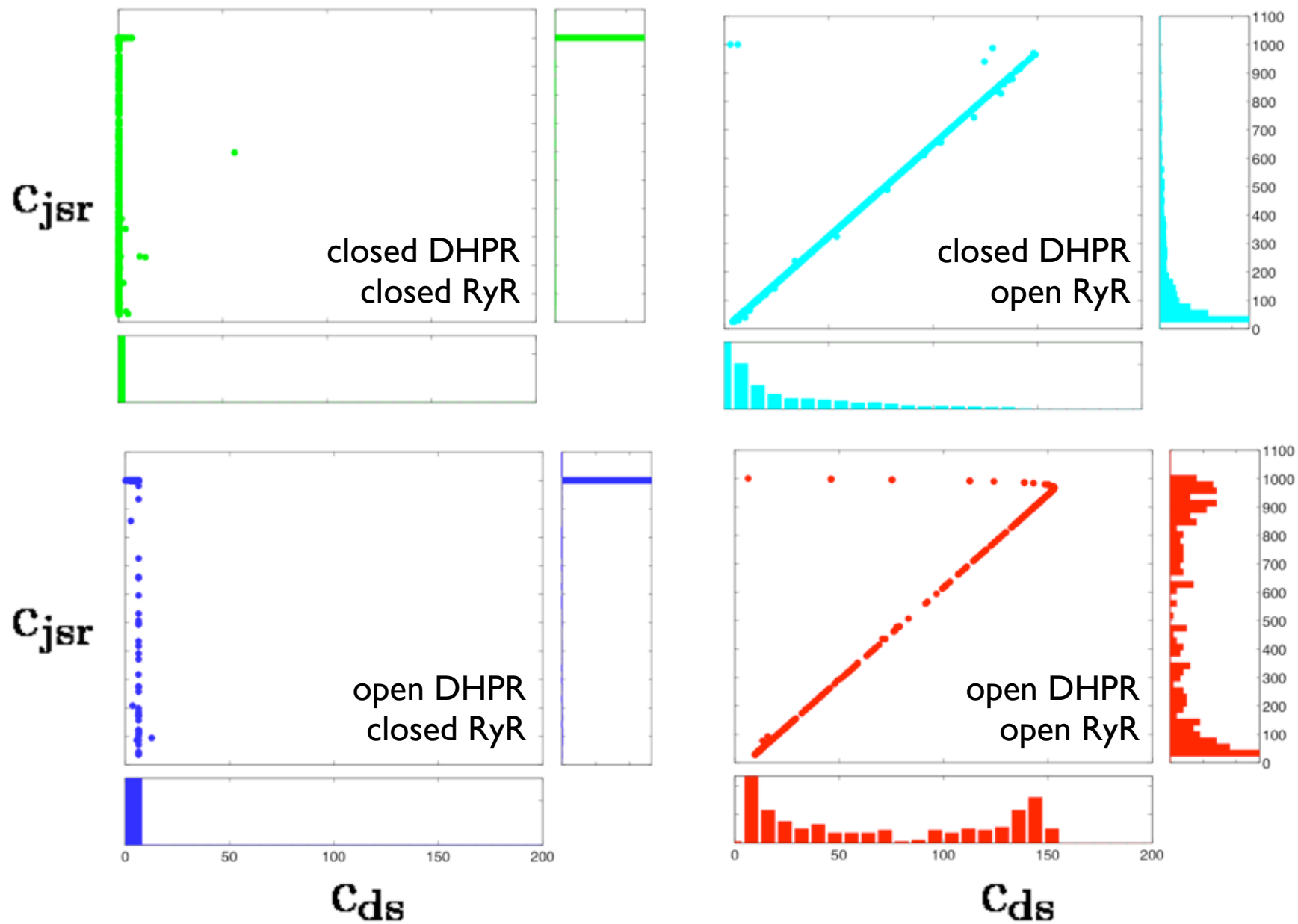


# Monte Carlo simulation of voltage-clamped cardiac myocyte

(1000 SFUs)

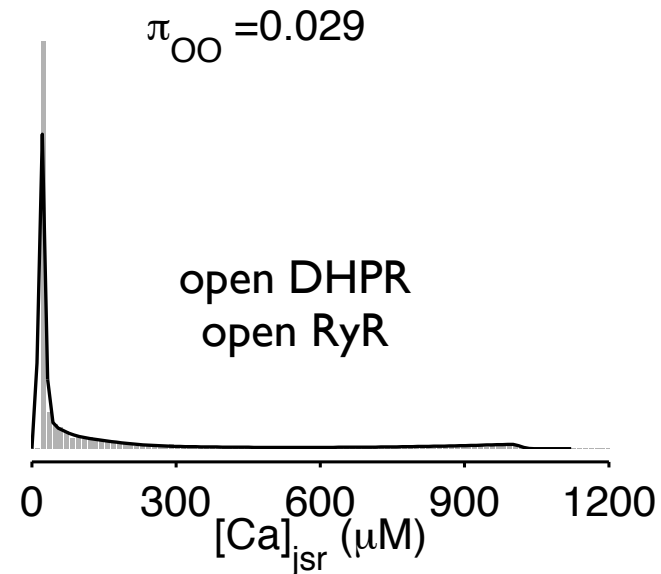
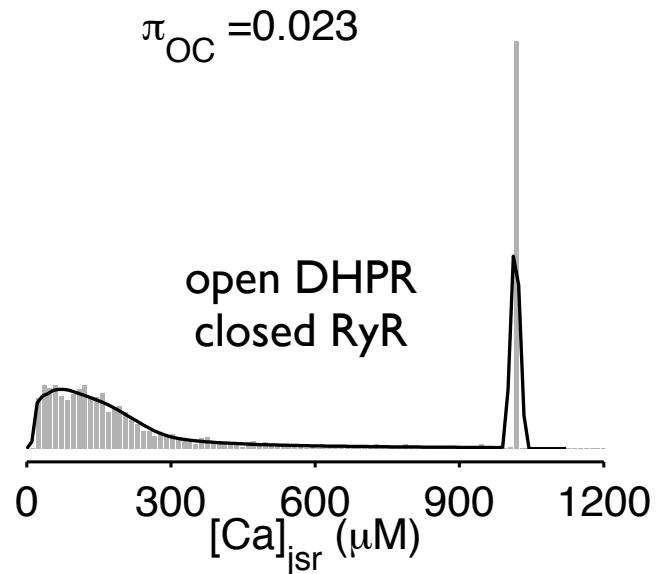
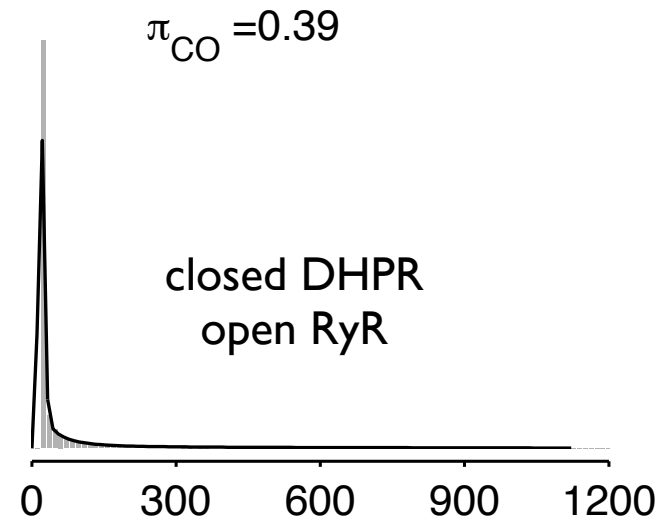
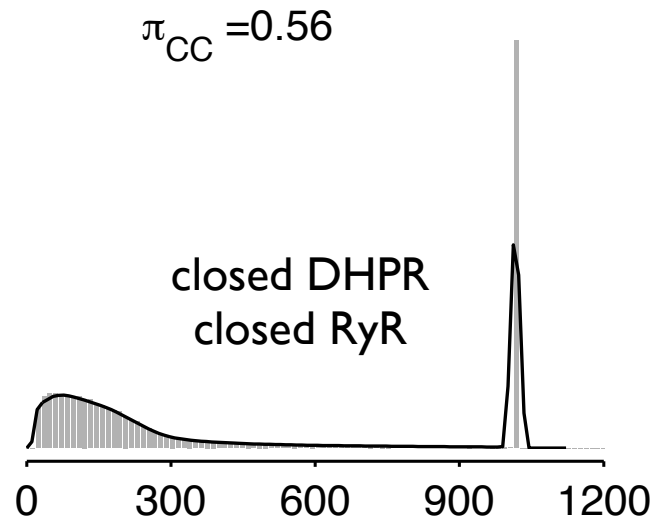


# Marginal densities show luminal depletion during voltage step

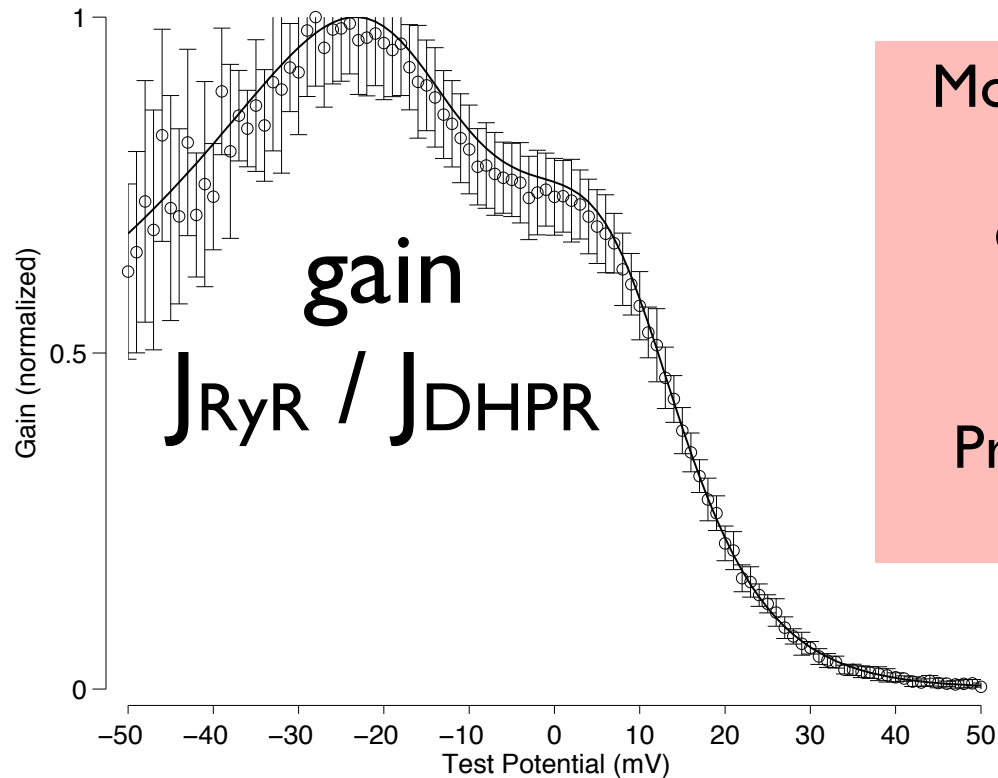
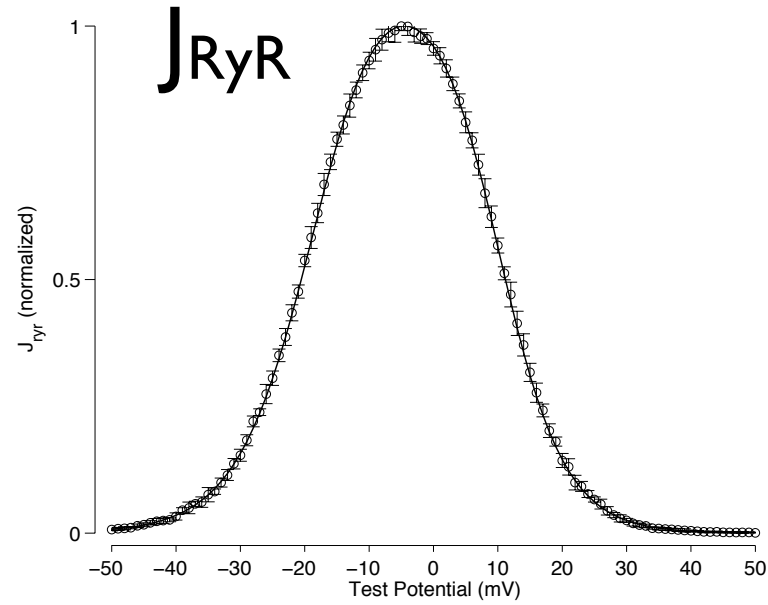
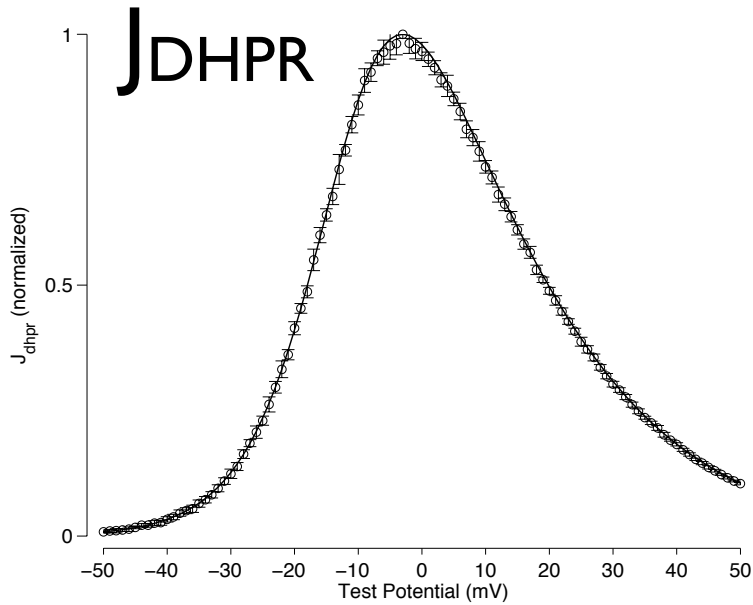


JSR is slow compared to diadic subspace...

Because JSR is slow compared to diadic subspace, we can reduce to one-dimensional densities (in terms of  $C_{jsr}$ )



# Probability density approach exhibits “gain and gradedness”



Monte Carlo using 1000 SFUs  
each point is 10 runs  
each run takes 3 minutes

Probability density approach  
each point takes 10 sec

# Computational efficiency of Monte Carlo and probability density approaches

Assuming the large N limit is of interest  
(20,000 SFUs  $\approx$  infinity)

Monte Carlo approach

discretization error  
time step ( $\Delta t$ )  
number of SFUs (N)

Probability density approach

discretization error  
time step ( $\Delta t$ )  
resolution of mesh ( $\Delta c$ )







# Acknowledgments



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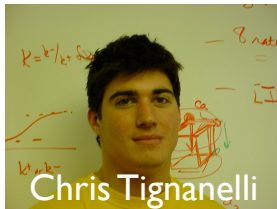
Jeff Groff



Blair Williams



Vien Nguyen



Chris Tignanelli

Longxiang Dai  
Joel Keizer  
Becky Lee  
Roy Mathias

Robert Miura  
Janet Oliver  
John Rinzel  
Artie Sherman  
Bridget Wilson  
Saleet Jafri  
Eric Sobie



MCB Signal Transduction



IBN Sensory Systems



Joint DMS/BIO/NIGMS Initiative  
to Support Mathematical Biology